Effect of 6 weeks' consumption of β-glucan-rich oat products on cholesterol levels in mildly hypercholesterolaemic overweight adults

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Abstract
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Keywords
oat, rich, glucan, β, consumption, weeks, 6, effect, adults, levels, overweight, hypercholesterolaemic, products, cholesterol, mildly

Disciplines
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Abstract
Several regulatory bodies have approved a health claim on the cholesterol-lowering effects of oat β-glucan at levels of 3–0 g/d. The present study aimed to test whether 1.5 g/d β-glucan provided as ready-to-eat oat flakes was as effective in lowering cholesterol as 3-0 g/d from oats porridge. A 6-week randomised controlled trial was conducted in eighty-seven mildly hypercholesterolaemic (≥5 mmol/l and <7.5 mmol/l) men and women assigned to one of three diet arms (25% energy (E%) protein; 45% E% carbohydrate; 30% E% fat, at energy requirements for weight maintenance): (1) minimal β-glucan (control); (2) low-dose oat β-glucan (1.5 g β-glucan; oats low - OL) or (3) higher dose oat β-glucan (3-0 g β-glucan; oats high - OH). Changes in total cholesterol and LDL-cholesterol (LDL-C) from baseline were assessed using a linear mixed model and repeated-measures ANOVA, adjusted for weight change. Total cholesterol reduced significantly in all groups (−7.8 (so 13.8)%), −7.2 (so 12.4)% and −5.5 (so 9.3)% in the OL, OL and control groups), as did LDL-C (−8.4 (so 18.5)%), −8.5 (so 18.5)% and −5.5 (so 12.4)% in the OL, OL and control groups), but between-group differences were not significant. In responders only (n 60), β-glucan groups had higher reductions in LDL-C (−18.3 (so 11.1)% and −18.1 (so 9.2)% in the OH and OL groups) compared with controls (−11.7 (so 7.9)%; P=0.044). Intakes of oat β-glucan were as effective at doses of 1.5 g/d compared with 3 g/d when provided in different food formats that delivered similar amounts of soluble β-glucan.

Key words: Cholesterol; Oats; β-Glucans; Solubility; Dosage

The viscous soluble fibre found in oats ((1→3),(1→4) β-glucan) has been demonstrated to have cholesterol-lowering effects(1,2). However, despite the majority of trials showing a cholesterol-lowering effect in hypercholesterolaemic subjects, no clear dose–response relationship has been demonstrated.

In addition, not all oat products show similar effects(3–5). The cholesterol-lowering properties appear to be linked to the physico-chemical properties of the soluble β-glucan fraction, rather than the total soluble fibre content per se(6). Putative effects have been attributed to an influence on the sequestration of bile acids in the gut, reducing re-absorption and return to the liver for further synthesis(7–9). The direct effects of oat bran on cholesterol levels might also be better seen in the immediate postprandial period through a dramatic effect on decreased chylomicron cholesterol(10). The cholesterol-lowering effects of β-glucan may be influenced by a number of factors, primarily the molecular weight, solubility and viscosity in the product (as consumed) and these are dependent on the food microstructure, dosage and the type of food processing that has been undertaken. For example, the process of enrichment may affect efficacy(11). Possible unfavourable structural changes that occur to β-glucan during commercial extraction include depolymerisation of the linear structure which decreases molecular weight and viscosity(12). Under mild extraction conditions, endogenous β-glucanase enzymes may not be deactivated and thereby further increase depolymerisation(12) which could lower efficacy. Endogenous β-glucanase enzymes are also active during food preparation(13), causing degradation of β-glucan in food products. In addition, freezing and storage may reduce the extractability of β-glucan in the intestine(14). On the other hand, processes in which oats are heated in the presence of water, such as baking, boiling and extraction(15,16), increase the solubility of β-glucan and viscosity which has been associated with an increase in bioactivity(6,17).

Abbreviations: E%, percentage of energy; LDL-C, LDL-cholesterol; OH, oats high; OL, oats low; RTE, ready-to-eat.

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Variation is in part explained by the physico-chemical properties of the β-glucan in the food and the diet overall[138]. Thus, the level of β-glucan administered is not the only factor to consider when evaluating the science.

In 1997, the US Food and Drug Administration[199] approved health claims for oat fibre, based on the relationship demonstrated between dietary soluble β-glucan fibre and a decrease in serum cholesterol concentrations. The Food and Drug Administration has concluded that fibre is efficacious in lowering total cholesterol and LDL-cholesterol (LDL-C) by about 5–10%, at doses of 3 g/d from either oat bran or rolled oats. This amount is provided by approximately 55 g oat bran (minimum 5.5% β-glucan) or 75 g rolled oats (4% β-glucan). More recently, the European Food Safety Authority[230], French Agency for Food, Environment and Occupational Health & Safety[231], the Joint Health Claims Initiative in the UK and the Food Directorate, Health Products and Food Branch of Health Canada have permitted claims for cholesterol-lowering effects at this level of β-glucan intake[232].

The present study aimed to assess the ability of the different doses of oat β-glucan delivered in different food formats (rolled oat porridge, a cereal bar and ready-to-eat (RTE) oat flake breakfast cereal) to lower cholesterol in mildly hypercholesterolaemic Australian adults.

Methods

Participants

The present study was a 6-week parallel, randomised, controlled, single-blind trial conducted with three arms. The two intervention groups were provided with oat porridge and oat-based cereal bars (group oats high (OH), higher β-glucan, 3.2 g/d) and RTE oat flakes and puffed rice and wheat bars (group oats low (OL), lower β-glucan, 1.5 g/d), respectively. The control group (minimal β-glucan) received cornflakes, puffed rice and wheat bars in plain packs.

Volunteers for the study were recruited via advertisements in the local media, including newspaper, television and radio. Approaches were also made to local medical practices, ambulance and fire services and via university staff emails. Inclusion criteria were as follows: men 25–75 years and premenopausal women >25 years or 5 years postmenopause (but not on hormone replacement therapy) with total serum cholesterol ≥5 and <7.5 mmol/l, BMI >20 and <32 kg/m²; regular breakfast cereal consumer (four to five times/week); stable weight (within 3 kg over past 3–6 months); and of general good health. Exclusion criteria include the following: major illnesses (including diabetes mellitus, known CHD and chronic renal failure); lipid-lowering medication; familial hypercholesterolaemia; fasting glucose >5.6 mmol/l; total cholesterol >7.5 mmol/l; taking dietary supplements that could influence cholesterol (e.g. fish oil); pregnancy/lactation; food allergies or habits inhibiting compliance with the study design; literacy and inadequate conversational English.

The present study was conducted according to the guidelines laid down in the Declaration of Helsinki and all procedures involving human subjects/patients were approved by the Human Research Ethics Committee of the University of Wollongong (HEIR/136). Written informed consent was obtained from all participants.

Baseline period

After eligibility had been determined on the basis of a telephonic interview and a fasting screening blood test to assess total cholesterol concentrations, eligible participants completed a computerised 7-d diet-history interview (DietAdvice, Xyris Software, 2009; Highgate Hill, QLD, Australia) to assess habitual nutrient intake. Dietary data were analysed using the FoodWorks software system (Xyris Software, version 5, 2007). Nutrient intake data were analysed in terms of energy and macronutrients using the AusNut (Allfoods) Revision 18 database. At 1 week later, eligible participants were randomly assigned to one of three diet groups and attended the university clinic for a dietary counselling session with a dietitian.

Randomisation and concealment

Randomisation was performed by a researcher independent of the subject interface using the RALLOC command in STATA (version 10.1; College Station, TX, USA). The procedure was stratified by sex and used permuted blocks, with the size and order of the blocks being random. All test foods were packaged in plain, opaque coded wrapping at the point of manufacturing and factory packaging. Since dietary advice was given in terms of food groups, it was not possible to blind the dietitians, but the participants were not informed as to their diet group allocation.

Interventions

Test foods

The between-group dietary difference occurred with the provision of alternative cereal foods in plain individual portion packs, batch packaged and provided by Cereal Partners Worldwide Limited (Rutherford, VIC, Australia). Participants were required to consume one packet of cereal and one cereal bar per d with a minimum of five of each per week. β-Glucan analyses (AOAC (Association of official Analytical Chemists) 995.16) of the products were provided by Medallion Labs, Minneapolis, MN, USA. Cereal bars contained 1.99% β-glucan; RTE oat flakes, 2.32% β-glucan; and oat porridge, 4.17% β-glucan. Control foods (RTE cornflakes and puffed rice bars with no β-glucan) were not analysed as maize and rice do not contain significant quantities of β-glucan. The amount of β-glucan provided in the product servings is shown in Table 1. The OH group received 3.24 g β-glucan/d and the OL group received 1.45 g β-glucan/d. The viscosity, solubility and molecular weight of all test foods were analysed by the Guelph Food Research Centre, Agriculture and Agri-Food, Canada. β-Glucan was extracted from the food samples (10 g dry weight basis) using conditions similar to those found in the upper gastrointestinal system. The extraction was done on 10 g of food at 37°C with digestive enzymes. The physiological extraction and characterisation of soluble β-glucans were carried out as described previously[233].
Table 1. Physico-chemical properties of test foods
(Mean values and standard deviations)

<table>
<thead>
<tr>
<th>Treatments ...</th>
<th>OH</th>
<th>OL</th>
<th>RTE oat flakes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Test foods</td>
<td>Rolled oats</td>
<td>Cereal bar</td>
<td></td>
</tr>
<tr>
<td>Serving size (g)</td>
<td>Mean</td>
<td>60</td>
<td>Mean</td>
</tr>
<tr>
<td>β-glucan content (%)</td>
<td>4.17</td>
<td>0.07</td>
<td>1.99</td>
</tr>
<tr>
<td>Dose (g β-glucan/d)</td>
<td>2.52</td>
<td></td>
<td>0.72</td>
</tr>
<tr>
<td>Extract viscosity (mPa s at 32 per second)*</td>
<td>16.03</td>
<td>0.48</td>
<td>3.80</td>
</tr>
<tr>
<td>Solubility (% total β-glucan)</td>
<td>35.93</td>
<td>0.35</td>
<td>24.09</td>
</tr>
<tr>
<td>C (g soluble β-glucan/d)</td>
<td>0.91</td>
<td></td>
<td>0.17</td>
</tr>
<tr>
<td>Mₚ (g/mol)</td>
<td>210000</td>
<td>23000</td>
<td>2400000</td>
</tr>
<tr>
<td>Mₚ x C (kg/mol x g/d)</td>
<td>1900</td>
<td></td>
<td>400</td>
</tr>
</tbody>
</table>

OH, oats high; OL, oats low; RTE, ready-to-eat; C, daily dose of soluble β-glucan; Mₚ, peak molecular weight.

* Viscosity of physiological extract from 10 g cereal.

The viscosity of the extract was measured using a rheometer with a cone-and-plate configuration. The solubility of β-glucan was calculated from the total β-glucan and the amount solubilised during the extraction process. The molecular weight of β-glucan in solution was analysed using size-exclusion HPLC. Measurements were done on duplicate extractions.

Dietary prescription. Each group was advised by Accredited Practising Dietitians on a healthy low-fat diet utilising the core food groups outlined in the Australian Guide to Healthy Eating^{24}. Energy intakes were calculated to equate to individuals’ estimated energy requirements for weight maintenance using the Mifflin equation^{25} and applying a physical activity factor of 1.25. Cereals and cereal bars provided were modelled into the dietary prescription to achieve macronutrient intakes of approximately 45–50% carbohydrates, 20–25% energy (E%) protein and 25–30 E% fat (Table 2). Participants were provided with educational material and eating plans. Participants were instructed not to eat products which would otherwise alter cholesterol such as those containing plant sterols and fish oil n-3 supplements while in the study and were given details of such foods.

Dietitians monitored participants throughout the study at follow-up appointments at 3 and 6 weeks.

Participants were issued with a diet compliance booklet in which they noted how many serves from each food group were consumed daily and completed a checklist of the number of packets of cereal or study bars consumed. Diet compliance booklets were returned at completion of the study.

Table 2. Baseline characteristics of the study subjects
(Mean values and standard deviations)

<table>
<thead>
<tr>
<th>Variables</th>
<th>OH (n = 30)</th>
<th>OL (n = 28)</th>
<th>Control (n = 32)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>Mean</td>
<td>52.43</td>
<td>51.93</td>
</tr>
<tr>
<td>Height (m)</td>
<td>Mean</td>
<td>1.69</td>
<td>1.69</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>Mean</td>
<td>77.00</td>
<td>77.87</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>Mean</td>
<td>26.74</td>
<td>27.28</td>
</tr>
<tr>
<td>Body fat (%)</td>
<td>Mean</td>
<td>30.08</td>
<td>32.10</td>
</tr>
<tr>
<td>Waist (cm)</td>
<td>Mean</td>
<td>93.90</td>
<td>92.91</td>
</tr>
<tr>
<td>Hip (cm)</td>
<td>Mean</td>
<td>104.43</td>
<td>103.63</td>
</tr>
<tr>
<td>SBP (mmHg)</td>
<td>Mean</td>
<td>128.76</td>
<td>132.12</td>
</tr>
<tr>
<td>DBP (mmHg)</td>
<td>Mean</td>
<td>76.59</td>
<td>77.07</td>
</tr>
<tr>
<td>Cholesterol (mmol/l)</td>
<td>Mean</td>
<td>5.97</td>
<td>6.12</td>
</tr>
<tr>
<td>TAG (mmol/l)</td>
<td>Mean</td>
<td>1.37</td>
<td>1.53</td>
</tr>
<tr>
<td>HDL (mmol/l)</td>
<td>Mean</td>
<td>1.53</td>
<td>1.61</td>
</tr>
<tr>
<td>Cholesterol/HDL ratio</td>
<td>Mean</td>
<td>4.08</td>
<td>4.28</td>
</tr>
<tr>
<td>LDL (mmol/l)</td>
<td>Mean</td>
<td>3.82</td>
<td>3.64</td>
</tr>
<tr>
<td>Glucose (mmol/l)</td>
<td>Mean</td>
<td>4.85</td>
<td>4.96</td>
</tr>
<tr>
<td>Insulin (pmol/l)</td>
<td>Mean</td>
<td>61.9</td>
<td>70.6</td>
</tr>
</tbody>
</table>

OH, oats high; OL, oats low; SBP, systolic blood pressure; DBP, diastolic blood pressure.
pressure readings were taken using a Dinamap XL Vital Signs Monitor (GE Healthcare, Chalfont St Giles, Buckinghamshire, UK). Body weight and percentage of body fat were measured while standing using bioelectrical impedance scales (Tanita BF-622W, Tanita Corporation of America, Arlington Heights, IL, USA). These scales have been validated and are thought to be a reasonable comparison with dual-energy X-ray absorptiometry as a reference method.\(^{(20)}\)

Activity and physical discomfort questionnaires were completed at the 3- and 6-week visits. Symptoms were scored on a scale from 1 to 6, with 1 being no symptoms and 2 to 6 being symptoms of increasing frequency/severity.

**Sample size**

Sample size was determined using an ANOVA approach, based on outcomes reported in a 6-week trial that used a similar dosage of oat-derived β-glucan.\(^{(27)}\) It was assumed that no change would occur in cholesterol in the control group, with the two intervention groups expected to show a change of 10% from baseline concentrations, with standard deviations of 10%. Using a values of significance of 0.05–0.01 and 90% power, a sample size of between nineteen and twenty-seven participants per group was required. To account for dropout, thirty participants per group were targeted for recruitment.

**Statistical analysis**

The primary outcome measures were fasting (10 h) total and LDL-cholesterol. Secondary outcomes were changes in fasting serum HDL, TAG, glucose, insulin and systolic blood pressure. Statistical analysis was performed using SPSS (version 17.0; SPSS Inc., Chicago, IL, USA). Model assumptions were checked before analysis. The analysis of primary and secondary variables was conducted using a linear mixed model for repeated measures or a general linear model for repeated measures as stated in the text. As weight loss is known to improve lipid, blood pressure, glucose and insulin measures, and weight decreased significantly during the study period in all groups, the analysis of all biochemical variables was adjusted for weight, as a continuous time-varying covariate. Adjusted and unadjusted analyses are presented. (The advantage of the use of the linear mixed model in this analysis is that partial datasets were incorporated and in this way, data from all subjects who commenced the study were included in the analysis.) An exploratory post hoc analysis was also conducted to investigate the effects in the responders. Responders were defined as those subjects who experienced a decrease of any magnitude (< 0.000 mmol/L) in LDL-C (n = 60). Non-responders were those who experienced no change or an increase (≥ 0.000 mmol/L) in LDL-C (n = 27). The analysis was also conducted in the form of a percentage change from baseline, defined as the difference between the values for week 6 and baseline, expressed as a percentage of the baseline measure. Factors that predicted LDL-responders were investigated using logistic regression analyses. Both univariate and multivariate analyses were conducted in the model building procedure. The following variables were considered in the logistic regression: study group, sex, age, baseline BMI, baseline percentage of body fat, waist circumference, baseline cholesterol, TAG and HDL. Multivariate analysis was conducted using an entry model (where all variables were included) and a backward elimination procedure with exclusion based on the likelihood ratio test. One-way ANOVA or t tests were conducted for the responder analysis with post hoc follow-up of significant results as indicated. Proportions were compared using Pearson’s χ² analysis. Results are presented as means and standard deviations unless otherwise indicated.

**Results**

**Participants**

Recruitment was conducted continuously over a 13-month period. A total of 328 people expressed interest in the present study and 127 were eligible for the confirmatory blood test, of whom ninety-five (fifty-one females, forty-four males) met the study criteria. Among these, five participants withdrew (four females, one male) before receiving the study foods. Randomisation was conducted for the remaining ninety participants by a researcher independent of the clinical interface (M. J. B.). The malefemale composition of each group was 15:15 in the OH group, 11:17 in the OL group and 15:17 in the control group. Withdrawals after the first counselling visit left 15:15 in the OH group (n = 30), 11:15 in the OL group (n = 26) and 15:16 in the control group (n = 31; Fig. 1).

The study sample was middle-aged (51 (sd 10.22) years), overweight (BMI 27.26 (sd 4.10) kg/m²) men and women with mildly elevated cholesterol levels but otherwise healthy (Table 2). There were no differences between the groups at baseline in the variables measured (P = 0.423–0.954).

**Clinical outcomes**

All three diet groups produced a reduction in total cholesterol levels (time effect P < 0.001), but there was no difference in reduction between the groups (Table 3 and Fig. 2; group effect: P = 0.565; interaction effect: P = 0.665). Similarly, all groups showed reductions in LDL-C (time effect P < 0.001) and HDL-cholesterol (time effect: P < 0.001), but between-group differences were not evident (Fig. 2). No change in TAG was evident (time effect: P = 0.279). Since there was no difference in response according to β-glucan dosage, the OH and OL groups were combined and a two-group exploratory analysis was conducted. Percentage reduction from baseline in LDL-C was −8.42 (sd 17.41) v. −5.48 (sd 12.36)% for the OH + OL and control groups, respectively (P = 0.363), resulting in a mean difference from baseline between the two groups of −2.94 (sd 3.22)%.

**Subgroup responder analysis**

Subgroup analyses of responders (n = 60) identified a trend towards a greater reduction in LDL-C from baseline at 6 weeks in the OH and OL groups (P = 0.086, one-way
ANOVA; Table 4). This difference was significant \( (P=0.044) \), one-way ANOVA) when LDL-C reduction was expressed as % change from baseline \((-18.3 \text{ (sd 11.1)\%}, -18.1 \text{ (sd 9.2)\%} \) and \(-11.7 \text{ (sd 7.9)\%}\) in the OH, OL and control groups, respectively), although the post hoc analysis adjusted for multiple comparisons showed that these between-group differences were of borderline significance for the two oat groups vs. controls \( (P=0.067 \text{ and } P=0.097 \text{ for the OH and OL groups, respectively; Tukey's honestly significant difference test). Post hoc analyses found no difference in response between the OH and OL groups; therefore, a two-group analysis (OH and OL groups combined) was performed. Between-group differences from baseline were significant for LDL-C \((-18.2 \text{ (sd 10.1)\%}, -11.7 \text{ (sd 7.9)\%}\); mean difference \(-28 \text{ (sd 2.3)\%}\) for the OH + OL vs. control groups, respectively; \( P=0.008 \).}

**Predictors of LDL-responders**

Baseline total cholesterol was the only significant predictor of LDL-responders, both in univariate and multivariate (backward stepwise) logistic regression models; the other variables entered (age, sex, BMI, body fat, TAG and HDL-cholesterol) were not significant. For every one unit increase in total cholesterol, there was a 2.850 times increase in the odds of being a responder \( (P=0.026) \). That is, increasing cholesterol is more likely to result in LDL response.

**Other outcomes**

There were no significant changes in glucose, insulin or blood pressure measurements. Homeostatic model assessment (HOMA) scores showed no significant difference for time,
Table 3. Changes in clinical parameters at 0, 3 and 6 weeks
(Mean values and standard deviations)

<table>
<thead>
<tr>
<th>Variable</th>
<th>OH (n 30)</th>
<th>OL (n 28)</th>
<th>Control (n 32)</th>
<th>OH (n 30)</th>
<th>OL (n 26)</th>
<th>Control (n 31)</th>
<th>OH (n 30)</th>
<th>OL (n 26)</th>
<th>Control (n 31)</th>
<th>Time</th>
<th>Group</th>
<th>Interaction</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total cholesterol (mmol/l)</td>
<td>5.97 ± 0.55</td>
<td>6.12 ± 0.54</td>
<td>6.03 ± 0.58</td>
<td>5.40 ± 0.87</td>
<td>5.40 ± 0.69</td>
<td>5.52 ± 0.67</td>
<td>5.49 ± 0.80</td>
<td>5.68 ± 0.77</td>
<td>5.67 ± 0.68</td>
<td>0.000 0.691 0.684</td>
<td></td>
<td></td>
</tr>
<tr>
<td>TAG (mmol/l)</td>
<td>1.37 ± 0.59</td>
<td>1.53 ± 0.73</td>
<td>1.56 ± 0.70</td>
<td>1.23 ± 0.44</td>
<td>1.54 ± 0.80</td>
<td>1.48 ± 0.81</td>
<td>1.34 ± 0.60</td>
<td>1.56 ± 0.58</td>
<td>1.55 ± 0.78</td>
<td>0.279* 0.306* 0.873*</td>
<td></td>
<td></td>
</tr>
<tr>
<td>HDL (mmol/l)</td>
<td>1.63 ± 0.31</td>
<td>1.61 ± 0.56</td>
<td>1.45 ± 0.37</td>
<td>1.43 ± 0.32</td>
<td>1.46 ± 0.55</td>
<td>1.32 ± 0.35</td>
<td>1.42 ± 0.31</td>
<td>1.48 ± 0.52</td>
<td>1.36 ± 0.37</td>
<td>0.000* 0.614* 0.589*</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cholesterol:LDL ratio</td>
<td>4.06 ± 0.83</td>
<td>4.26 ± 1.51</td>
<td>4.36 ± 1.01</td>
<td>3.88 ± 0.72</td>
<td>4.09 ± 1.37</td>
<td>4.43 ± 1.14</td>
<td>3.99 ± 0.79</td>
<td>4.15 ± 1.12</td>
<td>4.45 ± 1.15</td>
<td>0.286 0.284 0.355</td>
<td></td>
<td></td>
</tr>
<tr>
<td>LDL (mmol/l)</td>
<td>3.92 ± 0.55</td>
<td>3.84 ± 0.67</td>
<td>3.86 ± 0.55</td>
<td>3.40 ± 0.77</td>
<td>3.24 ± 0.70</td>
<td>3.52 ± 0.57</td>
<td>3.46 ± 0.69</td>
<td>3.49 ± 0.70</td>
<td>3.60 ± 0.53</td>
<td>0.165* 0.565* 0.346*</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Glucose (mmol/l)</td>
<td>4.85 ± 0.46</td>
<td>4.96 ± 0.53</td>
<td>4.86 ± 0.38</td>
<td>4.91 ± 0.43</td>
<td>4.92 ± 0.54</td>
<td>4.80 ± 0.45</td>
<td>4.81 ± 0.51</td>
<td>4.97 ± 0.61</td>
<td>4.84 ± 0.32</td>
<td>0.717 0.734 0.916</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Insulin (pmol/l)</td>
<td>8.92 ± 3.91</td>
<td>10.16 ± 8.29</td>
<td>9.10 ± 4.86</td>
<td>6.61 ± 3.58</td>
<td>9.26 ± 8.69</td>
<td>7.38 ± 4.55</td>
<td>8.45 ± 7.43</td>
<td>8.31 ± 8.93</td>
<td>7.69 ± 4.46</td>
<td>0.634* 0.473* 0.906*</td>
<td></td>
<td></td>
</tr>
<tr>
<td>SBP (mmHg)</td>
<td>128.8 ± 20.7</td>
<td>132.1 ± 17.1</td>
<td>131.8 ± 20.4</td>
<td>124.5 ± 19.4</td>
<td>131.1 ± 17.8</td>
<td>123.3 ± 16.5</td>
<td>123.2 ± 17.8</td>
<td>127.4 ± 15.3</td>
<td>122.9 ± 16.3</td>
<td>0.000 0.643 0.431</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Weight</td>
<td>77.1 ± 10.7</td>
<td>77.9 ± 16.4</td>
<td>81.1 ± 14.2</td>
<td>76.7 ± 10.5</td>
<td>76.5 ± 16.1</td>
<td>80.3 ± 14.2</td>
<td>75.6 ± 10.6</td>
<td>74.0 ± 13.4</td>
<td>80.3 ± 14.4</td>
<td>0.000 0.436 0.132</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Physical activity (Baecie)</td>
<td>7.93 ± 1.24</td>
<td>8.13 ± 1.24</td>
<td>7.80 ± 1.11</td>
<td>8.05 ± 1.32</td>
<td>8.34 ± 1.26</td>
<td>7.93 ± 1.20</td>
<td>8.01 ± 1.32</td>
<td>8.34 ± 1.26</td>
<td>7.93 ± 1.20</td>
<td>0.181 0.576 0.969</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

OH: cats high; OL: cats lean; SBP: systolic blood pressure.
* Adjusted for weight change.
for weight maintenance, by providing low-fat healthy eating advice, the oral intake of participants changed. Participants reduced, limited or completely removed high-energy, low-nutrient snack foods (e.g., potato crisps, chocolate, pastries) and alcohol from their regular eating patterns. A shift in energy consumption may have occurred by removing and limiting these foods and beverages from their diets, even when they were replaced with other foods.

Compliance with consumption of the test and control foods was high. All groups consumed both the breakfast cereal and cereal bar products equally. Of the forty-two serves of each pack provided, the control group consumed the mean of 38 (SD 4) cereals and 37 (SD 5) bars, comparing well with the 39 (SD 3) and 39 (SD 2) in the OL group and 37 (SD 5) and 36 (SD 8) in the OH group. Based on returned test product packets, only three subjects had compliance of less than 50%, but these participants were not excluded as intention-to-treat analyses were performed. No linear relationship between compliance and LDL-C change was demonstrated (r = 0.149, P = 0.277). Participants were able to consume the 60 g serving sizes of oats-containing cereals with no reported difficulty.

Due to the low number of reported symptoms of rating 3 or higher, data were recorded as no symptoms or symptoms and compared between the groups using χ² analysis. There was no significant difference between the groups in the frequency of reported gastrointestinal symptoms on flatulence, bloating and diarrhoea (P = 0.442, 0.201 and 0.467, respectively). There were no reported adverse events. There were no significant changes in the levels of physical activity between or within the groups during the intervention (Table 3).

**Physico-chemical properties of test foods.** The laboratory results for the physico-chemical properties of each of the test foods are shown in Table 1. The control samples contained insignificantly levels of β-glucan. The physiological extraction procedure was performed on these products and the viscosity of the extract from the puffed rice bar was 1.23 mPa s and the cornflakes extract had a viscosity of 0.93 mPa s measured at 52 s⁻¹, which was close to the viscosity of water (0.72 mPa s at 37°C). The β-glucan-containing foods had higher extract viscosities. The viscosities of the extracts from

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**Table 4. Change in LDL-cholesterol (LDL-C), according to the total group and LDL-C responders only**

<table>
<thead>
<tr>
<th>Group</th>
<th>Change from baseline (mmol/l)</th>
<th>sd</th>
<th>P (one-way ANOVA)</th>
<th>n</th>
<th>Change from baseline (%)</th>
<th>sd</th>
<th>P (one-way ANOVA)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>LDL-C</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total group</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>OH</td>
<td>−0.360</td>
<td>0.730</td>
<td>0.692</td>
<td>30</td>
<td>−8.38</td>
<td>18.46</td>
<td>0.712</td>
</tr>
<tr>
<td>OL</td>
<td>−0.665</td>
<td>0.651</td>
<td>0.266</td>
<td>26</td>
<td>−8.47</td>
<td>16.49</td>
<td>0.001</td>
</tr>
<tr>
<td>Control</td>
<td>−0.242</td>
<td>0.482</td>
<td>0.31</td>
<td>31</td>
<td>−5.48</td>
<td>12.36</td>
<td>0.712</td>
</tr>
<tr>
<td>Responders</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>OH</td>
<td>−0.738</td>
<td>0.489</td>
<td>0.086</td>
<td>21</td>
<td>−18.27</td>
<td>11.10</td>
<td>0.004*</td>
</tr>
<tr>
<td>OL</td>
<td>−0.729</td>
<td>0.467</td>
<td>0.17</td>
<td>17</td>
<td>−18.12</td>
<td>9.17</td>
<td>0.112</td>
</tr>
<tr>
<td>Control</td>
<td>−0.473</td>
<td>0.355</td>
<td>0.22</td>
<td>22</td>
<td>−11.68</td>
<td>7.87</td>
<td>0.712</td>
</tr>
</tbody>
</table>

OH, oats high; OL, oats low.

*Post hoc analysis showed that the reduction in LDL-C was less in the control group than in the intervention groups; however, the differences were not significant when adjusted for multiple comparisons (P = 0.067 and 0.097 for the OH and OL groups, respectively; Tukey's honestly significant difference test).
Table 5. Reported energy and macronutrient intakes at 0, 3 and 6 weeks* 
(Mean values and standard deviations)

<table>
<thead>
<tr>
<th>Variable</th>
<th>Baseline</th>
<th>Control (n 17)</th>
<th>3 weeks</th>
<th>Control (n 16)</th>
<th>6 weeks</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>OH (n 22)</td>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
</tr>
<tr>
<td>Energy (kJ)</td>
<td>10589 ± 3428</td>
<td>10797 ± 670</td>
<td>14291 ± 7549</td>
<td>10501 ± 4160</td>
<td>8589 ± 3759</td>
<td>8991 ± 3041</td>
</tr>
<tr>
<td>Protein (g)</td>
<td>19.09 ± 3.64</td>
<td>18.80 ± 2.87</td>
<td>20.06 ± 5.53</td>
<td>20.35 ± 3.21</td>
<td>21.50 ± 2.95</td>
<td>23.00 ± 8.21</td>
</tr>
<tr>
<td>Total fat (g)</td>
<td>31.84 ± 9.90</td>
<td>30.35 ± 2.68</td>
<td>29.33 ± 8.56</td>
<td>27.04 ± 8.71</td>
<td>26.35 ± 5.70</td>
<td>20.60 ± 5.80</td>
</tr>
<tr>
<td>Saturated fat (g)</td>
<td>11.30 ± 5.11</td>
<td>10.11 ± 7.23</td>
<td>10.42 ± 3.45</td>
<td>8.42 ± 2.60</td>
<td>7.79 ± 1.72</td>
<td>7.53 ± 1.68</td>
</tr>
<tr>
<td>Polyunsaturated fat (g)</td>
<td>5.43 ± 1.39</td>
<td>5.94 ± 2.28</td>
<td>5.71 ± 1.42</td>
<td>4.79 ± 1.51</td>
<td>5.27 ± 1.80</td>
<td>5.84 ± 2.07</td>
</tr>
<tr>
<td>Monounsaturated fat (g)</td>
<td>11.24 ± 2.61</td>
<td>11.46 ± 2.77</td>
<td>11.08 ± 3.85</td>
<td>11.21 ± 3.70</td>
<td>10.42 ± 2.68</td>
<td>9.95 ± 3.03</td>
</tr>
<tr>
<td>CHO (g)</td>
<td>46.81 ± 7.11</td>
<td>47.52 ± 9.81</td>
<td>46.18 ± 7.40</td>
<td>49.19 ± 8.14</td>
<td>49.37 ± 7.41</td>
<td>47.88 ± 5.33</td>
</tr>
</tbody>
</table>

OH, oats high; OL, oats low; E%, percentage of energy; CHO, carbohydrate; P/S, polyunsaturated:saturated.
* Not all participants completed dietary assessments at the follow-up visits.

Discussion

For this sample of otherwise healthy overweight adults with mild hypercholesterolemia, we found a significant decrease in the control group in the present study. In contrast, the control group also showed a significant decrease in the present study. In the control group, we found a significant decrease in the control group in the present study.

The effect of the changes in diet on the cholesterol level was significantly lower in the control group compared to the intervention group. However, in the intervention group, the effect was significantly lower in the intervention group compared to the control group. The effect of the changes in diet on the cholesterol level was significantly lower in the intervention group compared to the control group.

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were at least 80% compliant with study product consumption and had no major protocol violations. Their intention-to-treat analysis failed to demonstrate significant differences between the groups, even with a large sample size of 174 participants; so again, it may be worth pursuing the characteristics of responders in future studies that aim to expose the effects of β-glucan delivered in various food forms and dosages.

A recent systematic review of the consistent association between oat consumption and cardiovascular risk factors also found one study that showed cholesterol reduction in all groups including the low-fat diet. In our case, the healthy background diet appeared to influence weight, bearing in mind that weight loss alone can improve lipid profiles. In addition, concurrent changes in dietary fat occurred in the sample, including the displacement of saturated fat, which may confound the relationship between increased fibre intake and blood cholesterol levels. In the study reported here, we did not show effects on TAG, but the lowered HDL-cholesterol produced by the sample was consistent with the lower reported intake of dietary fat (<3.0% of energy). A recent meta-analysis of 30 controlled feeding studies showed that both modified-fat (30-50% of energy) and low-fat (18-30%) diets lower LDL-C, but modified-fat diets tend to lower HDL-cholesterol less, and may produce greater reductions in TAG. Finally, in the present study, the lack of effects on the secondary outcomes of blood pressure, glucose, insulin and homoeostasis model assessment (HOMA) indices probably reflected the relatively short period of study and the normal baseline levels recorded.

The present results may also have been influenced by the food matrix of the intervention foods. When oat-derived β-glucan was incorporated into bread and cookies and provided at a level of 6 g/d for 4 weeks, no cholesterol-lowering effect was observed. The authors concluded that the food matrix or the food processing, or both, may be influential in limiting the cholesterol-lowering properties of oat bran. In particular, a high-molecular-weight β-glucan molecule can entangle to form viscous solutions and high solubility must be maintained to ensure this effect. Reduction in molecular weight or solubility will therefore effect viscosity and have follow-on effects on physiological functionality. It has previously been shown that the viscosity of the in vitro extract of oat-containing foods positively correlates with cholesterol reduction and also glycemic response, and the satiety biomarkers cholecystokinin and peptide YY. The viscosity of β-glucan solutions is dependent mainly on the concentration of β-glucan in solution and its molecular weight. In the present study, the OH and OL (3 g β-glucan and 1.5 g β-glucan, respectively) test foods had near-equal cholesterol-lowering effects; yet molecular weights, solubility and viscosities were variable. Using viscosity as a measure of bioactivity, it could be expected that in the present study, cholesterol reduction should have been greater in OH vs. OL as the viscosity of rolled oats was higher (16-63) than the extruded oat flakes. Similarly, the molecular weight of β-glucan was also higher in both the rolled oats and cereal bar (OH group: 210 000 vs. 240 000 g/mol, respectively) than in the extruded oat flakes (OL group: 760 000 g/mol). However, previous studies have shown that high and medium molecular weights of β-glucan such as those of the two groups in the present study can significantly reduce LDL-C and modulate the blood glucose response. It was also shown in the present study that the dose of solubilised β-glucan was comparable (1:32 in OL vs. 1:08 g in OH) across the two intervention groups, which may also explain the similar results observed. Nevertheless, the present findings in the study reported here are consistent with other studies demonstrating that extrusion to produce an oats-containing breakfast cereal may only slightly decrease molecular weight yet increase solubility of β-glucan, compared with regular oats that have higher molecular weight with poorer solubility.

To take into account the drop in viscosity caused by partial depolymerisation in the RTE oat flakes during processing, the parameter $M_n \times C$ was calculated, where $M_n$ is the molecular weight of the β-glucan in solution and $C$ is the daily dose of soluble β-glucan. Wolfe et al. suggest that $M_n \times C$ may be a more robust measure of β-glucan bioactivity than the viscosity of extracted β-glucan, because molecular weight and $C$ are relatively insensitive to extrinsic factors. The $M_n \times C$ value for the OH treatment was 1600, whereas it was 1000 in the OL treatment. The similarity in these values may explain the similar reduction in cholesterol achieved with the two treatments in the present study. For comparison, these values are not that dissimilar to the values found for treatments at concentrations of 3 g β-glucan/d of medium molecular weight ($M_n = 528 000$) (3M) and 4 g β-glucan/d of low molecular weight ($M_n = 211 000$) (4L) for extruded oat bran cereals studied previously.

Hence, the 3L cereal had an $M_n \times C$ value of 1500 and significantly lowered LDL-C by 4.7% (P=0.012; n 64). However, the 4L cereal produced a non-significant LDL-C lowering of 2.3% (P=0.205; n 63), with an $M_n \times C$ value of 840. This may be due to the low $M_n \times C$ value reflecting a potentially lower bioactivity of the β-glucan. It should also be considered that the larger number of participants in the oat bran cereal study may have increased the ability to distinguish differences between the treatment and control groups compared with the present study in which less than half that number of participants were randomised to the OH (3 g β-glucan) treatment group.

There are a number of potential limitations to the present study. The study may have been under-powered, since sample size was determined assuming that no change in serum cholesterol from baseline would occur in the control group and that weight would remain stable in all groups. Further, the expected magnitude of reduction in LDL-C of 10% in the two intervention groups appears to have been too optimistic. The finding from secondary analyses that a higher baseline total cholesterol was predictive of LDL response to β-glucan consumption in responders is consistent with previous studies, but regression to the mean may have been a confounder in this analysis.

In conclusion, favourable reductions in total and LDL-C in healthy diets with oat β-glucan are supported by strong mechanistic evidence of oat β-glucan on cholesterol levels. However, in the present study, the incorporation of β-glucan...
from oat foods into a healthy low-fat diet for 6 weeks did not lead to a further significant decrease in serum cholesterol compared with a low-fat diet alone. No effect of the intervention diets was observed regarding the secondary outcomes (glucose, insulin resistance and blood pressure). The findings from the present study suggest that a smaller quantity (1.5 g/d) of medium-molecular-weight oat β-glucan with high solubility may be equally as effective as a higher quantity (3 g/d) of high-molecular-weight β-glucan. Further examination of these factors is warranted to assist in determining the lowest effective doses of oat β-glucan within different food matrices that may be influential in effectively reducing cholesterol in moderately hypercholesterolaemic populations.

Acknowledgements

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References

Cholesterol-lowering effects of oat foods


