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# Role of electrostatic interactions in the retention of pharmaceutically active contaminants by a loose nanofiltration membrane

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1                   **Role of Electrostatic Interactions in the Retention of**  
2                   **Pharmaceutically Active Contaminants by a Loose**  
3                   **Nanofiltration Membrane**

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## 18 **Abstract**

19 The role of electrostatic interactions in the separation of pharmaceuticals by a loose  
20 nanofiltration (NF) membrane was examined. While retention of the non-ionizable  
21 pharmaceutical carbamazepine was relatively independent of the solution chemistry,  
22 retention of the ionizable pharmaceuticals sulfamethoxazole and ibuprofen was strongly  
23 influenced by the solution pH and ionic strength. This finding is consistent with previous  
24 results investigating the effects of solution pH and ionic strength on the retention of  
25 proteins and organic acids. Pharmaceutical retention increases dramatically as the  
26 compound transforms from a neutral to a negatively charged species when the solution  
27 pH increases above its  $pK_a$  value. In contrast, solution ionic strength suppresses the  
28 double layer or the Debye screening length and therefore reduces the effectiveness of  
29 electrostatic interaction as a major retention mechanism by the loose NF. However,  
30 because of the formation of a hydrated layer around the charged functional groups of the  
31 pharmaceuticals and the fact that at a sufficiently high ionic strength the Debye length  
32 approaches a relatively constant value, this reduction in retention is relatively small. As a  
33 result, even at comparatively elevated ionic strengths, retention of the negatively charged  
34 sulfamethoxazole and ibuprofen by the loose NF membrane is considerably high.

35 **Keywords:** nanofiltration, pharmaceutically active compounds, charge repulsion,  
36 retention mechanisms

## 37 **1 Introduction**

38 In recent years, there has been considerable research effort focusing on removing specific  
39 individual contaminants instead of the surrogate and often ill-defined water quality  
40 indicators. This paradigm shift is mainly driven by stricter environmental regulations and  
41 legislation, greater need to utilize non-traditional water resources including water  
42 reclamation and water recycling. Further, partial removal of such compounds in water and  
43 wastewater treatment, and particularly the availability of advanced treatment technologies  
44 such as advanced oxidation, carbon adsorption, and membrane filtration as well as hybrid  
45 processes have contributed to a greater interest in understanding removal mechanisms.  
46 The centre of attention amongst such contaminants is a group known as pharmaceutically

47 active compounds (PhACs), which has been a major concern due to their widespread  
48 occurrence of sub microgram per liter concentrations in the aquatic environment. Most  
49 pharmaceuticals are not fully assimilated and hence excreted after administration to  
50 humans or animals. They can resist biodegradation during conventional wastewater  
51 treatment processes to a considerable extent, depending on their physicochemical  
52 characteristics. Several pharmaceuticals such as carbamazepine are highly persistent and  
53 are not removed at all by wastewater treatment plants (WWTPs) [1, 2]. In other cases, the  
54 high degradation rate of the pharmaceuticals can be virtually offset by their continuous  
55 introduction into the environment.

56 Research to date has clearly indicated the abundance of PhACs in sub microgram per liter  
57 concentrations in secondary treated effluent, surface water, ground water, and in extreme  
58 cases even in drinking water [3-5]. Because pharmaceuticals are designed to be  
59 biologically active, they have the potential to induce chronic sublethal effects on living  
60 organisms and any of such adverse health effects can instill serious consequences. It is  
61 therefore essential to prevent contaminants such as PhACs from entering the aquatic  
62 environment [4], and particularly so prior to potable water recycling.

63 Solute separation in a nanoporous membrane filtration process is driven mostly by size  
64 exclusion (also known as steric interactions) and electrostatic interactions. The former is  
65 often described by the hydrodynamic model where porous membranes are presented as  
66 bundles of straight, narrow, cylindrical pores and steric interactions are taken into account  
67 to correct for the hindered convection and diffusion of uncharged solute within the  
68 membrane pores [6]. Using the extended Nernst-Planck equation to include the Donnan  
69 or dielectric exclusion due to interactions between the membrane charged surface and the  
70 charged solute, several models has been developed to describe the latter. Amongst the  
71 early versions of these, the space charge model proposed by Wang et al. [7] assumed ions  
72 as point charges and focused almost exclusively on the electrostatic interactions between  
73 solute and the membrane charged surface. Recently, a more rigorous description of the  
74 dielectric exclusion behavior has also been developed for both monovalent and  
75 multivalent salts by Bowen and Welfoot [8]. Because mineral salts were used as model  
76 solutes in most of these studies, a knowledge gap remains with respect to electrostatic

77 interactions between charged organic compounds, particularly trace organics, and the  
78 membrane charged surface. Over the last few decades, due to commercial interest in  
79 several organic compounds such as proteins and lactate salts, contributions of electrostatic  
80 interactions to the transport of these solutes in porous membrane systems have also been  
81 the subject of extended research. Several studies have shown, for example, that protein  
82 transport is a function of the membrane surface charge characteristics and protein  
83 retention increases significantly under conditions where the membrane and protein have  
84 the same charge due to increased electrostatic repulsion. Furthermore, there is substantial  
85 experimental evidence that these electrostatic interactions between proteins and the  
86 membrane surface are dependent of the solution chemistry, such as pH and ionic strength.  
87 For example, Burns and Zydney [9, 10] and Nakao *et al.* [11] evaluated the effect of  
88 solution pH on the passage of proteins through ultrafiltration membranes and found that  
89 the maximum passage was typically attained near the protein isoelectric point where the  
90 protein has no net electrical charge. Millesime *et al.* [12] reported a significant decrease  
91 in the retention of bovine serum albumin and lysozyme from as high as 100% to only  
92 35% and 10%, respectively, as the solution ionic strength increased to 1 M by adding  
93 NaCl to the feed solution. This is consistent with the results reported earlier by Pujar and  
94 Zydney [13]. Both groups attributed this behavior to the decrease in electrostatic  
95 repulsion at high ionic strength.

96 To date, most theoretical analyses on the effects of electrostatic interactions on organic  
97 solute transport in membrane filtration were carried out with proteins [9-11, 13] and to a  
98 lesser extent lactate salts as model solutes [14, 15]. There is currently a lack of  
99 information with respect to the influence of solution chemistry on the separation process  
100 of charged pharmaceuticals, particularly by loose NF membranes. Although the transport  
101 behavior of small organic compounds may follow some of the trends observed for that of  
102 proteins, there are several fundamental differences in their physicochemical  
103 characteristics. First, proteins are relatively large macromolecules. While a protein  
104 molecule can contain multiple charged moieties, small organic compounds such as  
105 pharmaceuticals typically consist of a single charged functional group. Consequently, the  
106 conformation and size of a macromolecule can vary considerably due to intra-molecular  
107 electrostatic interactions between its charged groups. On the other hand, given the much

108 smaller size of pharmaceuticals, the contribution of the hydrated layer may play a role in  
109 their passage through the membrane pores. It is also noteworthy that studies investigating  
110 the separation of proteins and lactate salts often employed a very high background  
111 electrolyte concentration of up to 1M, typical to that of an industrial application rather  
112 than the water recycling context.

113 Given the significant role that membrane filtration has taken on in the water industry [16],  
114 especially for water recycling, the use of nanofiltration (NF) membranes for the removal  
115 of trace organics has been intensively investigated [17-21]. None of these studies,  
116 however, have examined the role of electrostatic interactions in the removal of PhACs by  
117 loose NF membranes. The objective of this study is to examine the role of electrostatic  
118 interactions in the removal of pharmaceuticals by such a loose polymeric NF membrane.  
119 Both the membrane and the pharmaceuticals were characterized in detail. The membrane  
120 retention behavior was related to the physicochemical properties of the pharmaceuticals  
121 and the membranes as well as to the solution chemistry. Variation in solution chemistry  
122 involved pH, ionic strength, and presence of divalent cations. On the basis of the results,  
123 the role of electrostatic interactions in the nanofiltration of the selected pharmaceuticals  
124 was elucidated and discussed.

## 125 **2 Materials and Methods**

### 126 *2.1 Nanofiltration membrane*

127 Flat sheet samples of a loose thin film composite NF membrane — denoted TFC-SR2  
128 (Koch Membrane Systems, San Diego, CA) — were used in this investigation. The  
129 membrane consists of a thin polyamide skin layer on top of a microporous polysulfone  
130 support. This membrane was selected because of its low salt and high organic matter  
131 retention which makes it a very desirable membrane if desalination or hardness removal is  
132 not required. It was received as flat sheet sample and was stored dry at 4 °C.

### 133 *2.2 Pharmaceutically active contaminants (PhACs)*

134 Three common pharmaceuticals — sulfamethoxazole, carbamazepine, and ibuprofen —  
135 representing three different drug categories, were selected for this study. **Figure 1** depicts

136 the structures of these compounds. The compounds were purchased from Sigma-Aldrich  
137 (Saint Louis, MO). The purity of these chemicals was reported to be 99 % or higher.  
138 Sulfamethoxazole is an important member of the sulfonamide antibacterial category and  
139 is probably the most frequently used antibiotic; carbamazepine is one of the most widely  
140 used anti-epileptic drugs; and ibuprofen is a common anti-inflammatory agent. Because  
141 they belong to three different drug categories, these pharmaceuticals have quite distinctive  
142 functional groups (**Figure 1**). The pharmaceuticals were first dissolved in pure methanol  
143 to make up stock solutions of 1 g/L. The stock solutions were stored at  $< 4$  °C and were  
144 used within 1 month.

145 **[Figure 1]**

### 146 *2.3 Cross flow membrane filtration system and filtration protocol*

147 A laboratory-scale membrane filtration unit with a rectangular stainless steel crossflow  
148 cell (effective membrane area of 40 cm<sup>2</sup> and a channel height of 2 mm) was used for the  
149 experiments. The temperature of the test solution was controlled using a chiller/heater  
150 (Neslab RTE 7) equipped with a stainless steel heat exchanger coil. Permeate flow was  
151 measured by a digital flow meter (Optiflow 1000, Agilent Technologies, Palo Alto, CA)  
152 connected to a PC, and the cross flow/feed flow was monitored with a rotameter.

153 Prior to each experiment, the membrane was stabilized at 12 bar using DI water for  
154 approximately 16 hours until there was no further variation in permeate flux. The feed  
155 reservoir temperature was kept constant at  $20 \pm 0.1$  °C throughout the experiment. Both  
156 permeate and retentate were recirculated back to the feed reservoir. In all filtration  
157 experiments, the background electrolyte solution contained 20 mM NaCl and 1 mM  
158 NaHCO<sub>3</sub>, and, unless otherwise stated, the pH was kept at 8.

159 Prior to experimenting with pharmaceuticals, the DI water used for membrane  
160 compaction was replaced with 7 liters of fresh DI water. The cross flow velocity and  
161 permeate flux were adjusted to 30.4 cm/s and 15 μm/s ( $54 \text{ Lm}^{-2}\text{h}^{-1}$ ), respectively.  
162 Pharmaceuticals were then spiked into the feed reservoir to make up a concentration of

163 500 µg/L. Approximately 1.5 mL of feed and permeate samples were taken for analysis  
164 at specified time intervals.

165 For experiments with variable pH, the solution was adjusted to pH 10.5 by addition of a  
166 proper volume of 1 M NaOH. The pH was then incrementally dropped to 3.5 using  
167 stepwise additions 1 M HCl. For experiments with variable electrolyte concentration, the  
168 initial solution contained 1 mM NaHCO<sub>3</sub> and the pH was kept at 8. NaCl solution (1 M)  
169 and CaCl<sub>2</sub> (0.2 M) were then added to the feed reservoir to incrementally increase the  
170 electrolyte concentration as required. The system was equilibrated for 1 hour prior to  
171 sample collection at each pH or electrolyte concentration value. Observed retention is  
172 defined as  $R = 100 \times (1 - C_P/C_F)$ , where  $C_P$  and  $C_F$  are the permeate and the feed  
173 concentrations, respectively.

#### 174 *2.4 Analytical methods*

175 A Shimadzu HPLC system (Shimadzu, Kyoto, Japan) equipped with a Supelco Drug  
176 Discovery C-18 column and a UV detector was used to analyze pharmaceutical  
177 concentration in the feed and permeate samples. Detection wavelengths for  
178 sulfamethoxazole and carbamazepine were set at 280 nm, and for ibuprofen at 225 nm.  
179 DI water (buffered with 0.025 M KH<sub>2</sub>PO<sub>4</sub>) and acetonitrile were used as the mobile  
180 phase. The mobile phase was delivered at a constant flow rate of 1 mL/min with a  
181 gradient set in accordance with the chromatographic behavior of the respective analytes.  
182 Analysis was carried out immediately following the nanofiltration experiments.

### 183 **3 Results and discussion**

#### 184 *3.1 Membrane characteristics*

185 Characteristics of the TFC-SR2 membrane have been previously described. It was  
186 reported to be relatively hydrophilic with contact angle of approximately 20° measured  
187 using the sessile drop technique [22]. The membrane retains a small percentage of  
188 calcium while sodium retention is virtually negligible [22]. Pore size measurement  
189 following the procedure described in our previous publication [23] indicates that this  
190 membrane has a relatively open pore size, with an average pore radius determined of 0.64  
191 nm. Despite being a loose NF membrane, the TFC-SR2 membrane has a relatively high



192 natural organic matter removal of 70-85% [24], which is the characteristic of the SR  
193 series. High flux and high salt passage in combination with a moderate to high organic  
194 matter removal make the TFC-SR2 membrane particularly attractive for water recycling  
195 as well as surface water treatment. In such applications, salt removal is unnecessary and  
196 often undesirable due to the energy loss with the buildup of osmotic pressure and the  
197 production of a brine that requires further treatment and disposal.

198 As expected, sodium chloride retention (estimated by conductivity measurements) by the  
199 TFC-SR2 membrane is very small, yet the membrane attains a considerable negative  
200 charge at pH values above pH 5 (**Figure 2**). Below pH 5, the membrane zeta potential  
201 varies sharply as a function of pH, from slightly positive at pH 2.5 to  $-10$  mV at pH 5.  
202 This is due to the deprotonation or protonation of the membrane functional moieties,  
203 which in this case consist of predominantly carboxylic and amine groups [25]. It is  
204 noteworthy that the membrane zeta potential can provide a good indicative parameter to  
205 assess the membrane surface charge density [25, 26]. The membrane zeta potential or  
206 charge density does have a small but discernible influence on the retention of sodium  
207 chloride (**Figure 2**). This observation suggests that electrostatic interaction plays a small  
208 role in the separation process of ionic species by the TFC-SR2 membrane. Retention is  
209 smallest near the isoelectric point of the membrane. Due to the relatively large pore  
210 radius of the membrane (0.64 nm) with regard to the hydrated radii of the chloride or  
211 sodium ions, which have been reported to be 0.20 and 0.18 nm [27], respectively, the  
212 effect is relatively small. However, as will be discussed in a later section, electrostatic  
213 interactions can contribute substantially to the retention of charged organic molecules.

214 **[Figure 2]**

### 215 *3.2 Physicochemical properties of the selected PhACs*

216 Because of the differences in functional groups, the three pharmaceuticals selected for  
217 this study exhibit markedly different physicochemical properties (**Table 1**). While  
218 carbamazepine is uncharged at common pH conditions typical of natural water or  
219 wastewater, both ibuprofen and sulfamethoxazole exhibit a wide variation in speciation  
220 (or charge) and physicochemical properties. At pH below its  $pK_a$  value (pH 4.9),

221 ibuprofen is a neutral species. Above this  $pK_a$  value, ibuprofen attains a negative charge.  
222 Speciation of sulfamethoxazole as a function of pH has been described elsewhere [28].  
223 This pharmaceutical can exist in positive, neutral, as well as negative forms as it  
224 possesses two ionizable amine groups. At pH above the compound's second  $pK_a$  value  
225 (pH 5.7), sulfamethoxazole exists predominantly as a negatively charged species. It is  
226 noteworthy that values of the hydrophobicity presented in **Table 1** are assumed to  
227 represent characteristics of the compounds in their neutral form. Data for other pH values  
228 can be obtained by considering the effective partition coefficient (commonly known as  
229 logD) for the dissociative systems and can be found in databases such as SciFinder.  
230 Variations in charge and other physicochemical properties as a function of pH may have  
231 important implications for the separation mechanisms of these pharmaceuticals.

232 [Table 1]

### 233 *3.3 Steric hindrance and electrostatic interactions*

234 **Figure 3** presents the concentration of sulfamethoxazole, carbamazepine, and ibuprofen  
235 in both permeate and feed solutions as a function of time during filtration with the TFC-  
236 SR2 membrane at pH 4.0. Because of their low hydrophobicity, both sulfamethoxazole  
237 and carbamazepine do not adsorb to the membrane at these experimental conditions,  
238 which is evident from their constant feed concentrations for the duration of the  
239 experiments. Interestingly, despite the fact that the TFC-SR2 has a relatively hydrophilic  
240 surface, ibuprofen adsorbs considerably to this membrane, which manifests itself as  
241 decreasing feed concentration (from 500 to about 300  $\mu\text{g/L}$ ) until equilibrium is reached  
242 (**Figure 3**). In its neutral form, ibuprofen is a highly hydrophobic compound as reflected  
243 by its high  $\log K_{ow}$  value (**Table 1**) and this observed adsorption can probably be attributed  
244 to hydrophobic interactions between ibuprofen and hydrophobic domains within the  
245 membrane polymer matrix. It is noteworthy that the molecular size of ibuprofen (**Table**  
246 **1**) is considerably smaller than the average pore size of the TFC-SR2 membrane, and  
247 therefore, adsorption is not confined to the membrane surface and hence can take place  
248 throughout the polymer structure. As can be seen in **Figure 3**, this adsorption initially  
249 appears as a low permeate concentration (and hence high retention) while equilibrium is  
250 reached after about 2.5 hours of filtration.

251

[Figure 3]

252 At equilibrium when no further adsorption is observed, ibuprofen retention of  
253 approximately 35% can be inferred from **Figure 3**. This is significantly higher than the  
254 retentions of both sulfamethoxazole and carbamazepine, which are practically negligible  
255 despite the fact that these pharmaceuticals have about the same molecular size (**Table 1**).  
256 As discussed previously, at this experimental condition of pH 4, sulfamethoxazole and  
257 carbamazepine exist in their neutral form while 10% of ibuprofen still carries a negative  
258 charge. Consequently, this can be a major factor for the difference in retention between  
259 ibuprofen and the other two pharmaceuticals. Further reason for this can be due to the  
260 adsorption of the neutral ibuprofen onto the membrane and pore surfaces, leading to a  
261 higher observed retention, some of which may also be related to pore size reduction.  
262 Similar observation has also been made when the effects of adsorption on protein  
263 retention by ultrafiltration membranes was evaluated [29]. In addition, a sufficiently high  
264 dipole moment (above 3 D) can induce an electrostatic attraction between the membrane  
265 surface and the polar centers of the molecule [30]. Because the dipole moment of both  
266 sulfamethoxazole and carbamazepine is quite high (**Table 1**), the molecules tend to  
267 approach the membrane pore head on which results in a lower retention [30].

268 At pH 8, where both sulfamethoxazole and ibuprofen are negatively charged  
269 (carbamazepine remains neutral and hence has not been investigated), electrostatic  
270 attraction induced by the compound's polarity can be overcome by the electrostatic  
271 repulsion (**Figure 4**). Due to electrostatic repulsion (or charge exclusion),  
272 sulfamethoxazole is retained to some extent while the retention of ibuprofen is  
273 considerably higher than that at pH 4. It is noteworthy that at this pH, ibuprofen does not  
274 adsorb to the membrane as indicated by the constant feed and permeate concentrations  
275 throughout the experiment.

276

[Figure 4]

277 The effect of speciation (in other words the variation in charge of a species as a function  
278 of pH) on retention is further illustrated in **Figure 5**. Because carbamazepine is neutral at  
279 all pH values in examined here, carbamazepine retention is constant and independent of

280 solution pH (and membrane charge). In contrast, retention of both sulfamethoxazole and  
281 ibuprofen varies markedly, resembling their speciation curves as a function of pH, with  
282 the exception of high ibuprofen retention at low pH due to adsorption. This is consistent  
283 with several other studies where the nanofiltration of lactic or amino acids was  
284 investigated [15, 31-33].

285 **[Figure 5]**

286 The presented results indicate a distinctive difference between the retention behaviors of  
287 ionizable organic compounds and inorganic salts such as NaCl. As reported in an earlier  
288 section, sodium chloride retention was small and relatively constant over a wide pH range  
289 from 2 to 8 (**Figure 2**). Again, one can speculate that the retention behavior is attributed  
290 to the relative size difference between the solute and the membrane pore.  
291 Sulfamethoxazole and ibuprofen are considerably larger than the chloride ion (**Table 1**).

292 It is also interesting to point out that ibuprofen retention is consistently higher than that of  
293 sulfamethoxazole. This can possibly be explained by the fact that ibuprofen is an organic  
294 acid. Therefore, when dissociated, the negative ibuprofen species has a higher charge  
295 density than that of sulfamethoxazole, which is deprotonated via the dissociation of an  
296 amine group. Such a higher charge density would result in not only an increase in charge  
297 repulsion, but also a larger molecule hydrated size.

### 298 *3.4 Influence of background electrolyte: Monovalent Salt*

299 Solution ionic strength is directly related to the Debye length or the double layer thickness  
300 of the charged solutes and at the membrane surface, which in turn governs electrostatic  
301 interaction in NF processes. It is hence expected to influence the separation of charged  
302 solutes by NF membranes to some extent. Indeed, experimental data at pH 8 for the two  
303 charged compounds, presented in **Figure 6**, appear to strongly support this hypothesis.  
304 As ionic strength increases (represented by an increase in sodium chloride concentration),  
305 the Debye length becomes smaller. In other words, electrostatic interaction between the  
306 membrane and charged molecules is screened resulting in lower electrostatic repulsion  
307 and hence reduced retention. However, it should be noted that the Debye length is a

308 characteristic length for the range of electrostatic interaction and does not represent the  
309 actual dimension of a charged particle or surface. One can imagine the Debye length as  
310 an extension of a membrane pore (or more precisely a surface functional group) and of a  
311 molecule (or its charged functional group). One can further picture that this Debye length  
312 ‘diminishes’ the effective size of a pore or increases the effective size of a molecule. A  
313 qualitative picture of such an effect is shown in **Figure 7**. If dimensions are of the right  
314 proportions, that is if Debye length is of the same order of magnitude as the size  
315 difference between molecule and pore, then such a variation may be a determining factor  
316 in the retention of a charged molecule by a charged membrane. It is noted that Figure 7  
317 presents a conceptual model and does not show the Debye length overlap.

318 **[Figure 6]**

319 **[Figure 7]**

320 It can also be confirmed that this influence of ionic strength on retention is absent when  
321 the solute is neutral at pH 4 (**Figure 8**). Consequently, the molecule retention is constant  
322 as sodium chloride concentration in the feed increase up to 70 mM (or 4095 mg/L).  
323 While an increase in salt concentration can occur for varying feed waters, for most  
324 nanofiltration and reverse osmosis membranes that retain salt such an increase in salt  
325 concentration also occurs in the polarized layer near the membrane surface.

326 **[Figure 8]**

### 327 *3.5 Influence of background electrolyte: Divalent Salts*

328 As expected, the influence of  $\text{CaCl}_2$  concentration on the retention of negatively charged  
329 pharmaceuticals is more dramatic (**Figure 9**). Calcium is a divalent ion and thus is more  
330 effective in screening the molecule and membrane charge. Furthermore, calcium can also  
331 reduce the membrane charge because of its binding capacity to the membrane surface  
332 functional groups. As calcium chloride concentration increases to 8 mM, retention of the  
333 negatively charged pharmaceuticals appears to reach a plateau value while a concentration  
334 of 80 mM of NaCl is necessary for sulfamethoxazole and ibuprofen retention to reach a  
335 relatively constant value (**Figure 6**).

336

[Figure 9]

337 In both cases, this plateau retention value is significantly higher than the retention of  
338 neutral sulfamethoxazole and ibuprofen (**Figures 3 and 5**). This is possibly due to a  
339 limitation in the compressibility of the double layer at increasing ionic strength. As  
340 demonstrated in **Figure 7**, the Debye length decreases as the ionic strength or salt  
341 concentration increases following an exponential decay pattern. At concentration above  
342 80 mM for NaCl, an increase in ionic strength only results in a small incremental decrease  
343 in the Debye length. Similar conclusion can also be inferred for CaCl<sub>2</sub>, although in this  
344 case, CaCl<sub>2</sub> reduces the Debye length more effectively and therefore above 8 mM of  
345 CaCl<sub>2</sub>, the Debye length decrease is negligible as CaCl<sub>2</sub> concentration is further increased.

346 A further consideration is the formation of a hydrated layer around the negatively charged  
347 moiety of the pharmaceuticals and the membrane functional groups. While very little is  
348 known about the hydration of polymers and organic molecules, this effect possibly results  
349 in a considerable increase in their apparent size. For example, the thickness of a  
350 monolayer of water molecules is approximately 0.1 nm. Although the hydration energy  
351 or hydrated radius of the negatively charged pharmaceuticals investigated here are not  
352 available in the literature, it appears that the hydration energy of the anions is stronger  
353 than that of the cations [26]. Consequently, hydration may also be a considerable factor  
354 contributing to the difference in retention at a sufficiently high ionic strength between  
355 neutral and negatively charged pharmaceuticals as observed in **Figures 8 and 9**.

#### 356 **4 Conclusion**

357 Results reported here indicate that retention of the ionizable pharmaceuticals at the low  
358 concentrations examined is strongly influenced by solution pH and ionic strength. These  
359 results are consistent with previous studies on retention of proteins and organic acids such  
360 as lactic acid. Solution pH governs the speciation (or charge) behavior of the compound  
361 and therefore the retention mechanisms. Pharmaceutical retention increases dramatically  
362 as the compound transforms from a neutral to a negatively charged species as solution pH  
363 increases above its  $pK_a$  value. Ionic strength screens the molecule and membrane charges  
364 and therefore reduces the effectiveness of electrostatic repulsion as a major retention

365 mechanism by the loose NF membrane. However, such a reduction is relatively small and  
366 at a comparatively high ionic strength, retention of the negatively charged  
367 sulfamethoxazole and ibuprofen by the loose NF membrane remains considerably high at  
368 50-85%. This is probably attributed to the incompressibility of the Debye length at a  
369 sufficiently high ionic strength (about 80 mM) and the formation of a hydrated layer  
370 surround the negatively charged moieties of the pharmaceuticals.

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**Table 1.** Physicochemical Properties of Pharmaceuticals <sup>a</sup> [34], <sup>b</sup> [35], <sup>c</sup> [36], <sup>d</sup> Calculated by the Wilke-Chang and the Stokes-Einstein equations [37]. These values present the size of the neutral compounds, <sup>e</sup> estimated using HyperChem 7.0 [38], <sup>f</sup> [39], <sup>g</sup> [40].

Pharmaceutical	MW (g/mol)	$pK_a$	Stokes radius (nm)	Log $K_{ow}$	Dipole Moment (D)
Sulfamethoxazole	253.3	$pK_{a1} = 1.7^a$ $pK_{a2} = 5.6^a, 5.7^b$	0.38 <sup>d</sup>	0.89 <sup>c</sup>	5.4 <sup>e</sup>
Carbamazepine	236.3	2.3 <sup>c</sup>	0.37 <sup>d</sup>	2.45 <sup>c</sup>	3.6 <sup>e</sup>
Ibuprofen	206.3	4.4 <sup>e</sup> - 4.9 <sup>c</sup>	0.34 <sup>d</sup>	3.5 <sup>c</sup> , 4.13 <sup>f</sup>	1.8 <sup>g</sup>

498 FIGURE CAPTIONS

499 Figure 1. Chemical structures of the three pharmaceuticals used in this study.

500 Figure 2. Zeta potential of the TFC-SR2 membrane (in a background electrolyte solution  
501 containing 20 mM NaCl and 1 mM NaHCO<sub>3</sub>) and conductivity retention (feed solution  
502 contained 20 mM NaCl and 1 mM NaHCO<sub>3</sub>) by the TFC-SR2 membrane as a function of  
503 pH.

504 Figure 3. Feed and permeate concentration of the uncharged (a) sulfamethoxazole, (b)  
505 carbamazepine, and (c) ibuprofen species as a function of filtration time for the TFC-SR2  
506 membrane. The feed solution contained 500 µg/L of the corresponding pharmaceuticals  
507 in a background electrolyte solution containing 20 mM NaCl and 1 mM NaHCO<sub>3</sub>. Other  
508 experimental conditions were as follows: cross flow velocity = 30.4 cm/s, permeate flux =  
509 15 µm/s (54 Lm<sup>-1</sup>h<sup>-1</sup>), pH = 4, and temperature = 20 °C.

510 Figure 4. Feed and permeate concentration of the negatively charged (a)  
511 sulfamethoxazole and (c) ibuprofen species as a function of filtration time for the TFC-  
512 SR2 membrane. The feed solution contained 500 µg/L of the corresponding  
513 pharmaceuticals in a background electrolyte solution containing 20 mM NaCl and 1 mM  
514 NaHCO<sub>3</sub>. Other experimental conditions were as follows: cross flow velocity = 30.4  
515 cm/s, permeate flux = 15 µm/s (54 Lm<sup>-1</sup>h<sup>-1</sup>), pH = 8, and temperature = 20 °C.

516 Figure 5. Retention of sulfamethoxazole, carbamazepine, and ibuprofen by the TFC-SR2  
517 as a function of the solution pH. The feed solution contained 500 µg/L of the  
518 corresponding pharmaceuticals in a background electrolyte solution containing 20 mM  
519 NaCl and 1 mM NaHCO<sub>3</sub>. Other experimental conditions were as follows: cross flow  
520 velocity = 30.4 cm/s, permeate flux = 15 µm/s (54 Lm<sup>-1</sup>h<sup>-1</sup>), and temperature = 20 °C.

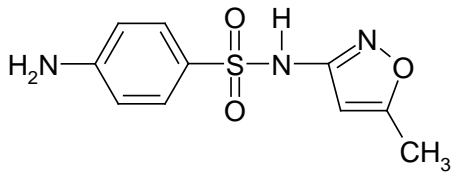
521 Figure 6. Retention of anionic sulfamethoxazole and ibuprofen by the TFC-SR2 as a  
522 function of the solution NaCl concentration. The feed solution contained 500 µg/L of the  
523 corresponding pharmaceuticals in a background electrolyte solution containing 1 mM  
524 NaHCO<sub>3</sub> and varied concentration of NaCl. Other experimental conditions were as

525 follows: cross flow velocity = 30.4 cm/s, permeate flux = 15  $\mu\text{m/s}$  ( $54 \text{ Lm}^{-1}\text{h}^{-1}$ ), pH = 8,  
526 and temperature = 20 °C.

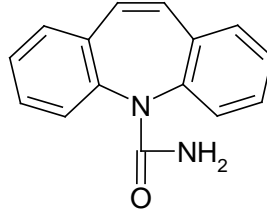
527 Figure 7. The calculated Debye length as a function of the solution NaCl concentration  
528 and a schematic description of the interplay between the Debye length of a charged  
529 molecule and an idealized membrane pore (a) relatively high retention at low ionic  
530 strength and (b) lower retention at high ionic strength.

531 Figure 8. Retention of the uncharged sulfamethoxazole and carbamazepine species by the  
532 TFC-SR2 as a function of the solution NaCl concentration. The feed solution contained  
533 500  $\mu\text{g/L}$  of the corresponding pharmaceuticals in a background electrolyte solution  
534 containing 1 mM  $\text{NaHCO}_3$  and varied concentration of NaCl. Other experimental  
535 conditions were as follows: cross flow velocity = 30.4 cm/s, permeate flux = 15  $\mu\text{m/s}$  ( $54$   
536  $\text{Lm}^{-1}\text{h}^{-1}$ ), pH = 4, and temperature = 20 °C.

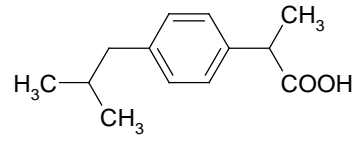
537 Figure 9. Retention of anionic sulfamethoxazole and ibuprofen by the TFC-SR2 as a  
538 function of the solution  $\text{CaCl}_2$  concentration. The feed solution contained 500  $\mu\text{g/L}$  of the  
539 corresponding pharmaceuticals in a background electrolyte solution containing 1 mM  
540  $\text{NaHCO}_3$  and varied concentration of  $\text{CaCl}_2$ . Other experimental conditions were as in  
541 Fig. 6.



Sulfamethoxazole



Carbamazepine



Ibuprofen

**Figure 1**

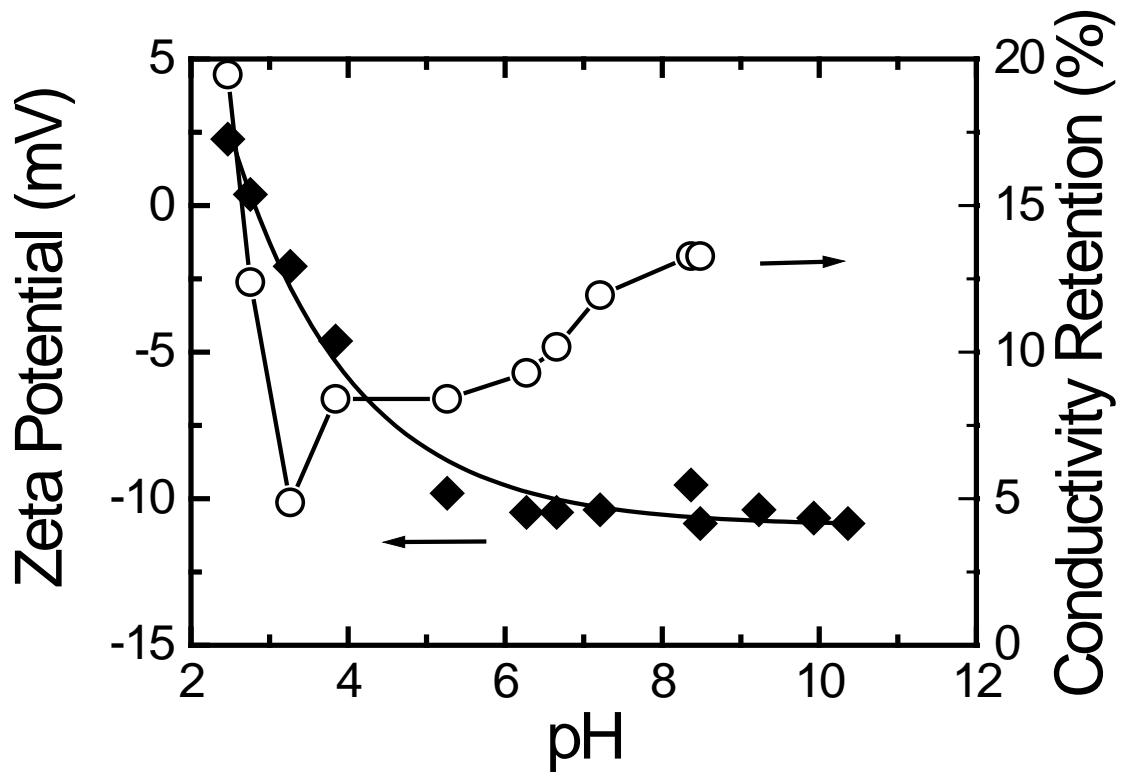


Figure 2

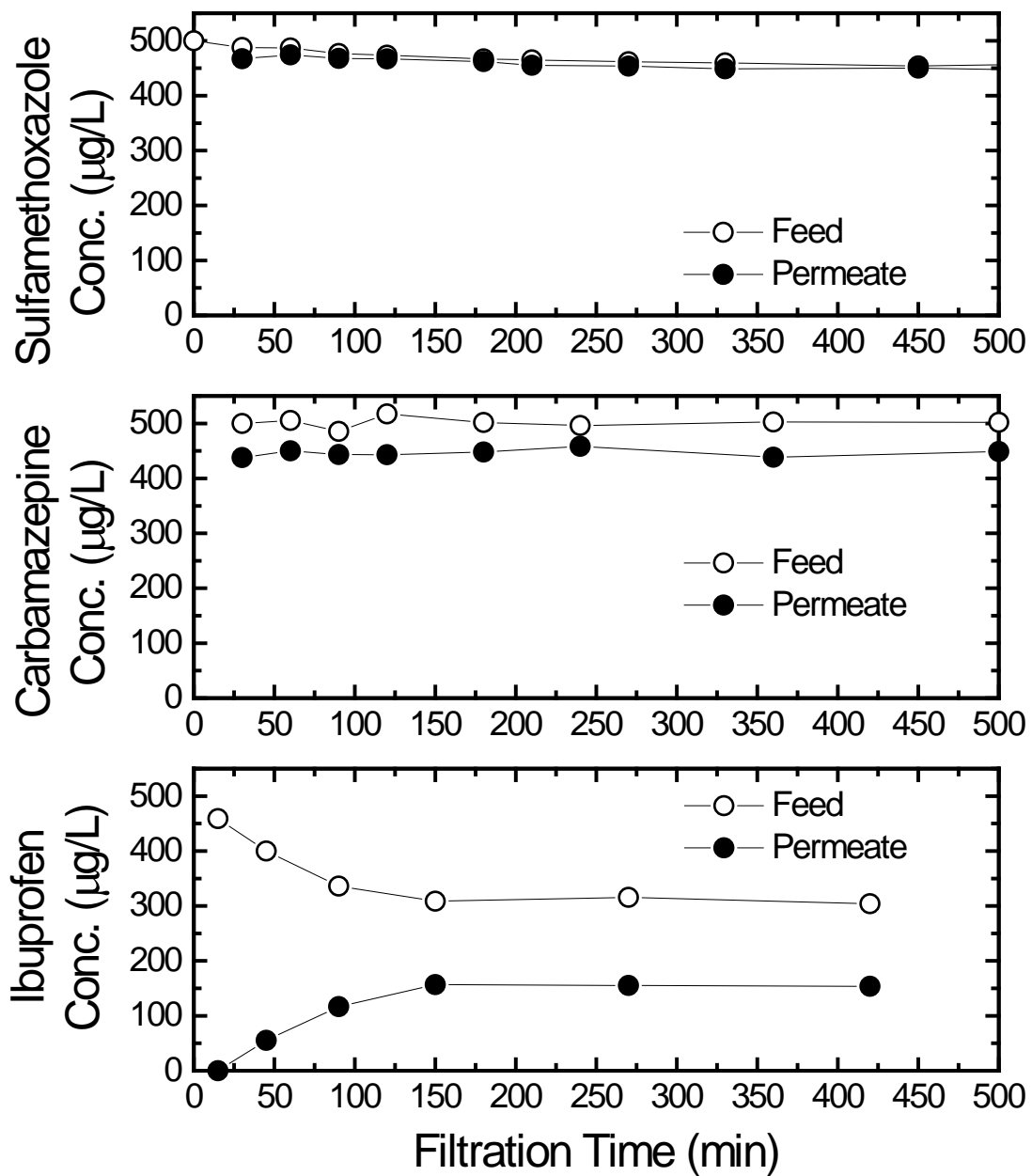


Figure 3



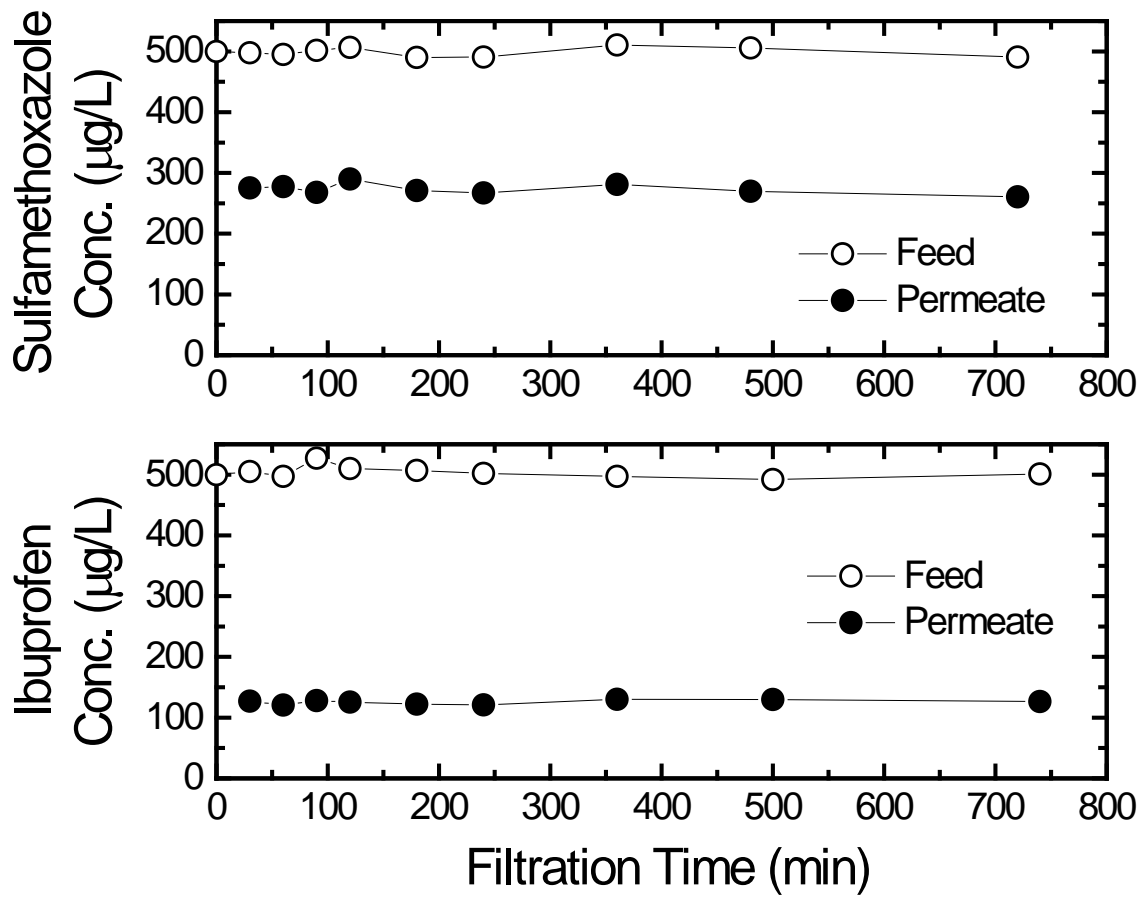


Figure 4

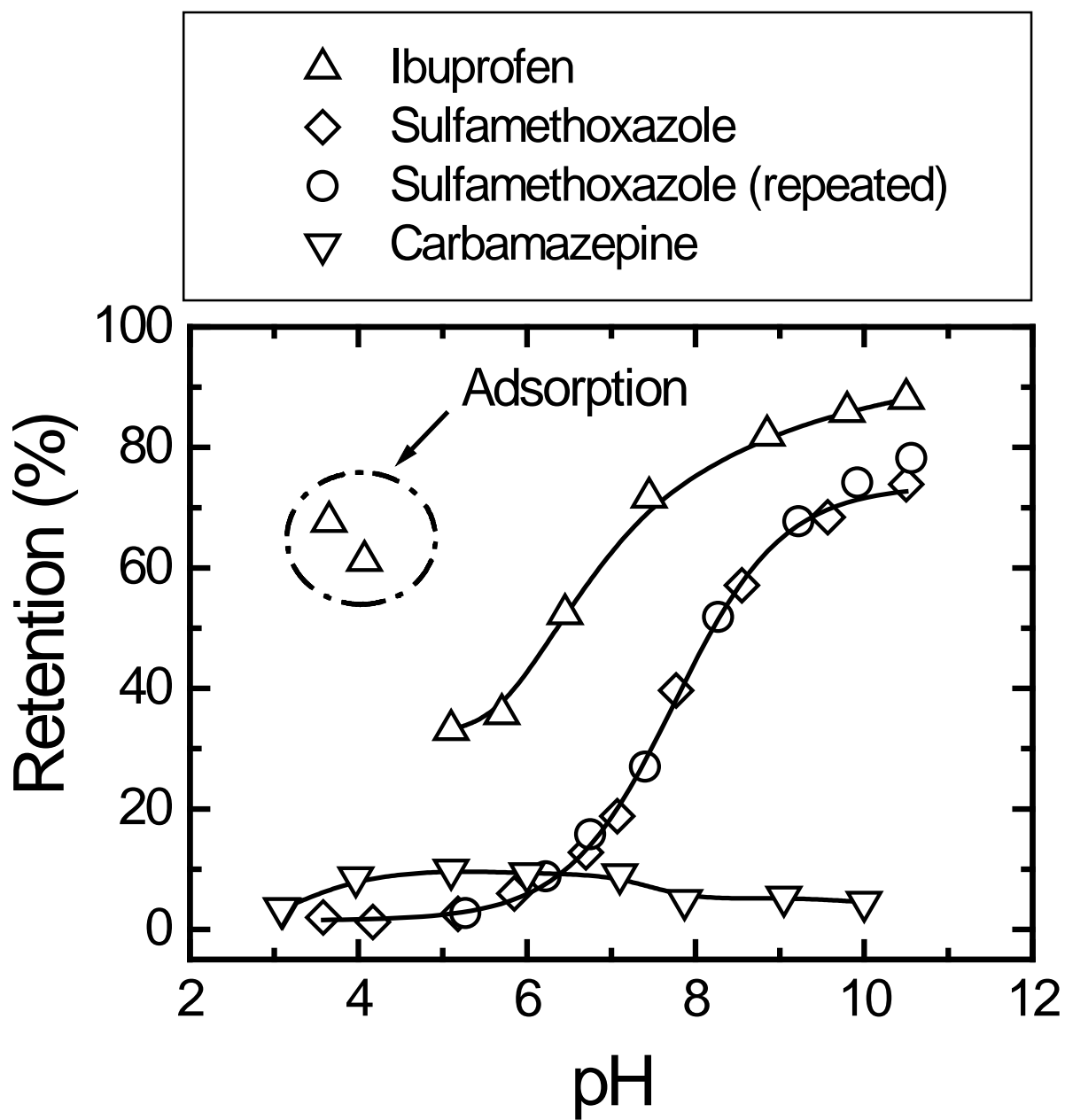


Figure 5

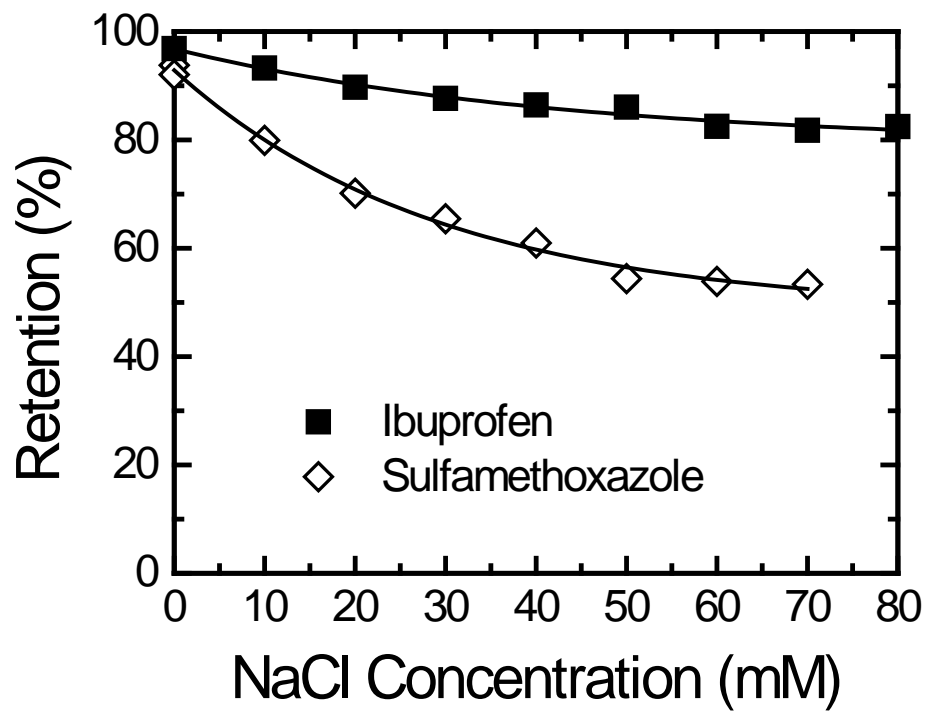
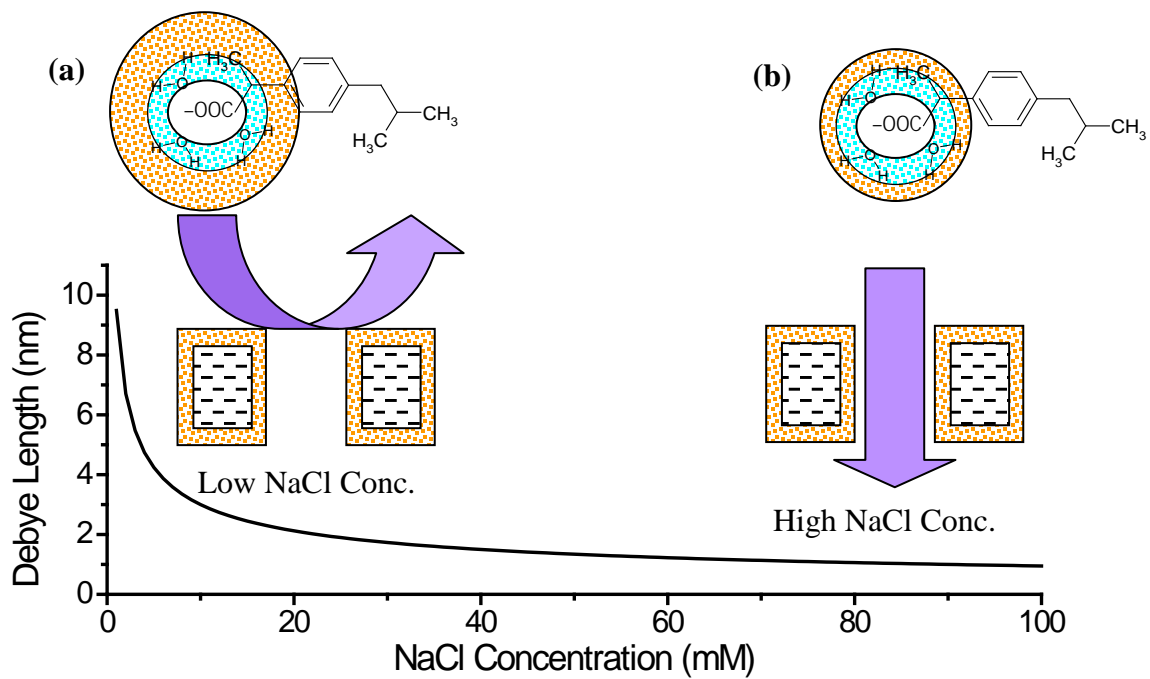


Figure 6



**Figure 7**

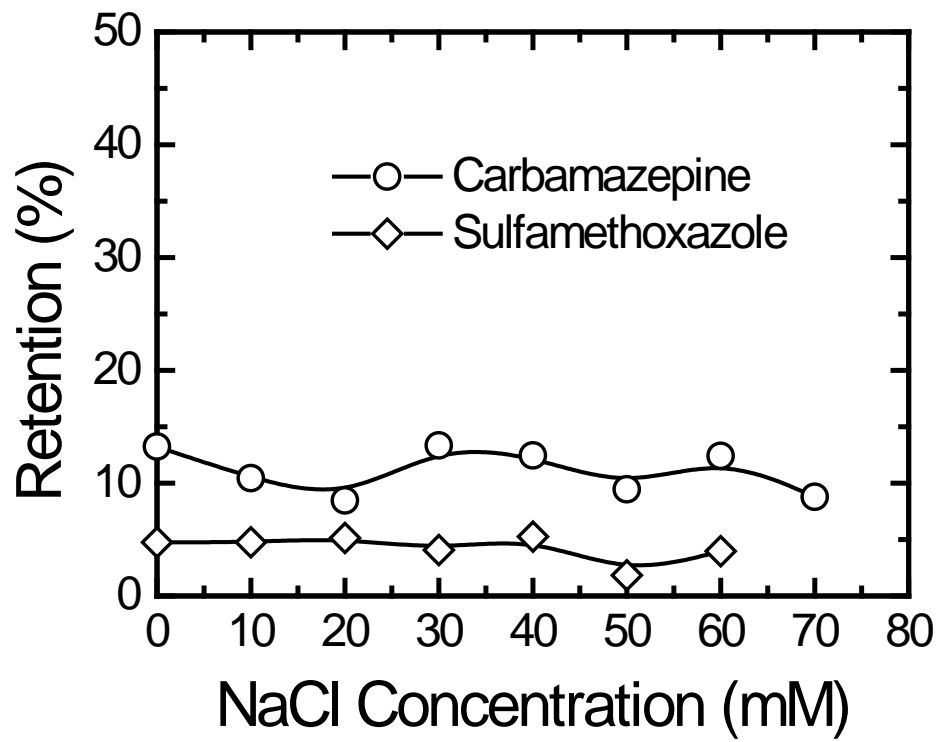


Figure 8

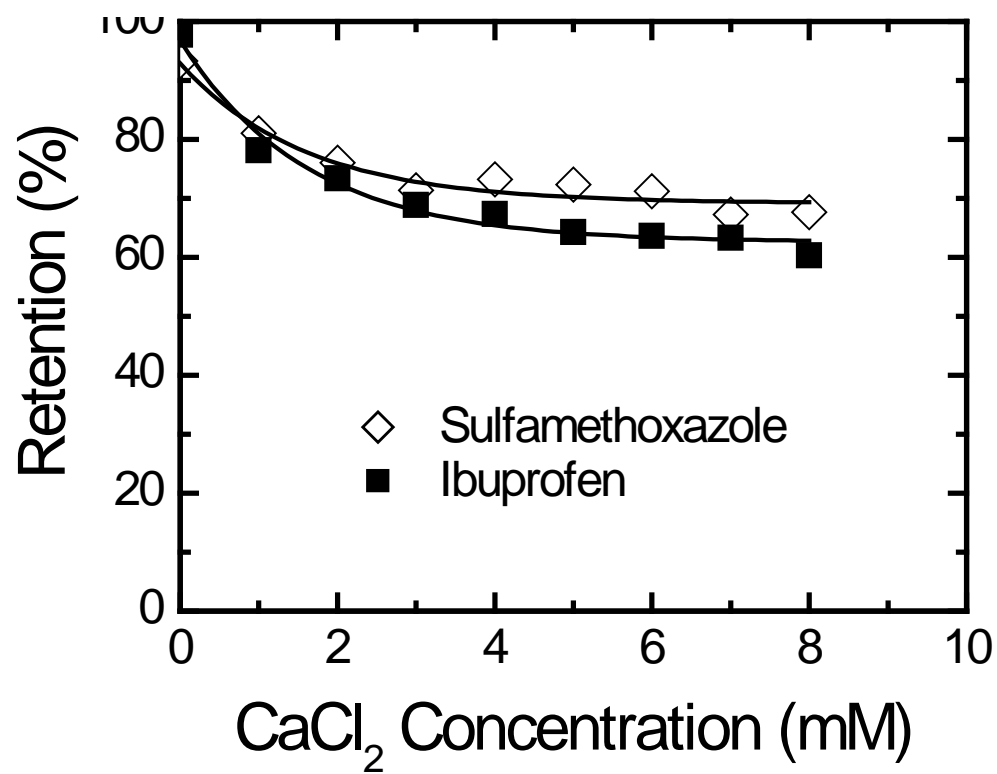


Figure 9