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Synthesis, characterisation and application of inherently conducting polymer nanoparticles

Orawan Ngamna University of Wollongong

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SYNTHESIS, CHARACTERISATION AND APPLICATION OF INHERENTLY CONDUCTING POLYMER NANOPARTICLES

A thesis submitted in fulfilment of the requirements for the award of the degree

DOCTOR OF PHILOSOPHY

from the

UNIVERSITY OF WOLLONGONG

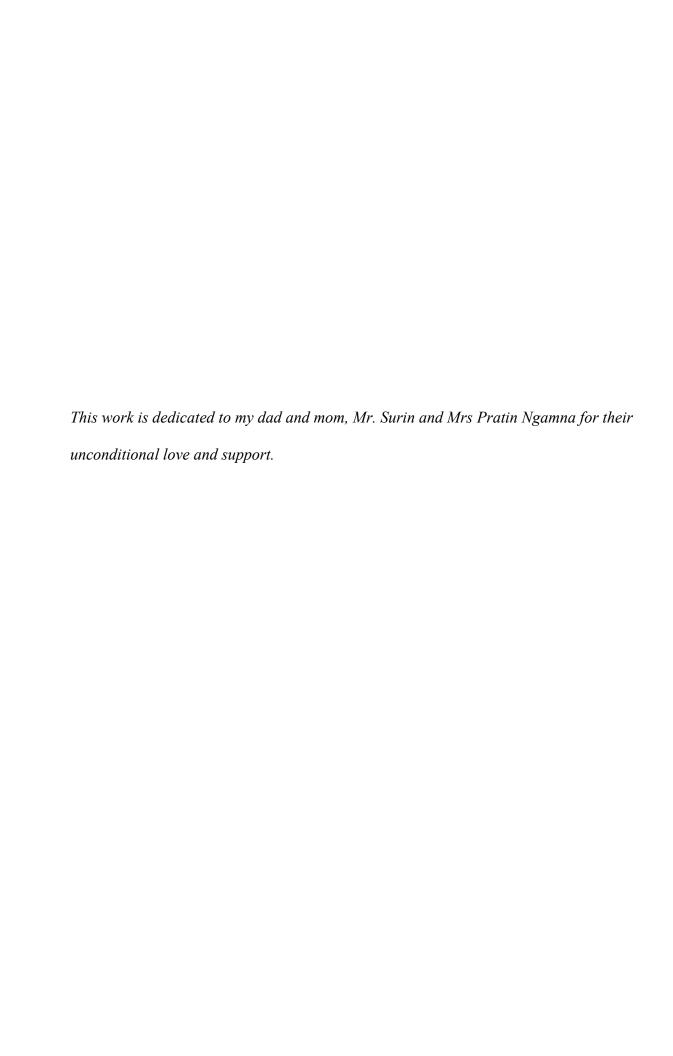
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ORAWAN NGAMNA, B.Sc.(Food Technology)

INTELLIGENT POLYMER RESEARCH INSTITUTE

DEPARTMENT OF CHEMISTRY

February 2006



CERTIFICATION

I, Orawan Ngamna, declare that this thesis, submitted in fulfilment of the requirements for the award of Doctor of Philosophy, in the Department of Chemistry, University of Wollongong, is wholly my own work unless otherwise referenced or acknowledged. The document has not been submitted for qualifications at any other academic institution.

Orawan Ngamna

February 2006

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- 1. S.E. Moulton*, P.C. Innis, L.A.P. Kane-Maguire, O. Ngamna, and G.G. Wallace, "Polymerisation and characterisation of conducting polyaniline nanoparticle dispersions", *Current Applied Physics*, 2004, **4** (2-4), 402-406.
- A. Morrin, O. Ngamna, A.J. Killard, S.E. Moulton, M.R. Smyth*, and G.G. Wallace*, "An Amperometric Enzyme Biosensor Fabricated from Polyaniline Nanoparticles", *Electroanalysis*, 2005, 17 (5-6), 423-430.
- 3. O. Ngamna, A. Morrin, S.E. Moulton, A.J. Killard, M.R. Smyth, and G.G. Wallace*, "An HRP based biosensor using sulphonated polyaniline", *Synthetic Metals*, 2005, **153** (1-3), 185-188.
- 4. A. Morrin, F. Wilbeer, O. Ngamna, S.E. Moulton, A.J. Killard, G.G. Wallace, and M.R. Smyth*, "Novel biosensor fabrication methodology based on processable conducting polyaniline nanoparticles", *Electrochemistry Communications*, 2005, 7 (3), 317-322.
- 5. J.M. Pringle*, O. Ngamna, J. Chen, G.G. Wallace, M. Forsyth and D.R. MacFarlane, "Conducting Polymer Nanoparticles Synthesized in an Ionic Liquid by Chemical Polymerisation", *Synthetic Metals*, in press.

6. O. Ngamna, S.E. Moulton and G.G. Wallace, "Incorporation of Dye into Conducting Polyaniline Nanoparticles", *Polymer*, accepted.

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ABBREVIATIONS

μ micro

 μ viscosity

 $\lambda \qquad \qquad wavelength$

*k*_B Boltzman's constant

γ surface tension

 Ω Ohm

v frequency

°C degree Celsius

 Θ angle

η refractive index

σ conductivity

 ρ resistivity

υ scan rate

A ampere

Å angstrom

A anion

 $A_{\rm i}$ acivity of species i

ABTS 2,2'-azino-bis(3-ethylbenzthiazoline-6-sulphonic acid)

AFM atomic force microscopy

Ag/AgCl silver/silver chloride reference electrode

Ag/Ag⁺ silver/silver ion reference electrode

APS ammonium peroxydisulphate

BSA bovine serum albumin

BFEE boron-fluoride ethyl ether

C Cunningham slip correction factor

ca. approximately

CB Carbolan Blue

CCD charge-coupled device

cm centrimeter

cm⁻¹ wave number

CMC critical micelle concentration

CV cyclic voltammetry

d diameter

Da dalton

D_{AB} diffusion coefficient

DBSA dodecylbenzene sulphonic acid

DLS dynamic light scattering

E potential

E⁰ standard reduction potential

EB emeraldine base

emiTFSA 1-ethyl-3-methylimidazolium-

bis(trifluoromethane-sulfonyl)amide

ES emeraldine salt

F Faraday constant

g gram

GC glassy carbon

HCSA camphorsulphonic acid

HPLC high performance liquid chromatography

HRP horseradish peroxidase

h hour

ICP inherently conducting polymer

IL ionic liquid

ITO indium tin oxide

k thousand

L litre

LB leucoemeraldine base

m metre

M molar

MAS 2-methoxyaniline-5-sulphonic acid

mg milligram

min minute

ml millilitre

mM millimolar

MoAb monoclonal antibody

mS millisiemen

Mw molecular weight

nm nanometre

OCP open circuit potential

PANI polyaniline

PB perniganiline base

PBS phosphate buffer saline

PEI poly(ethyleneimine)

PLL poly(L-lysine)

PMAS poly(2-methoxyaniline-5-sulphonic acid)

PPy polypyrrole

PS perniganiline salt

PSS poly(sodium 4-styrene sulfonate)

Pt platinum

PTh polythiophene

PTTh polyterthiophene

PVA poly(vinyl alcohol)

PVP poly(4-vinylpyridine)

PVS polyvinylsulphonate

Py pyrrole

r radius

RM rapid mixing

RMS root mean square

rpm round per minute

S siemen

SCE saturated calomel electrode

SDS sodium dodecylsulfate

SEM scanning electron microscopy

SHE standard hydrogen electrode

SPM scanning probe microscopy

T temperature

TBAP tetrabutylammonium perchlorate

TEM transmission electron microscopy

Th thiophene

TTh terthiophene

V volt

vs. versus

w/v weight by volume

w/w weight by weight

ABSTRACT

Synthesis of inherently conducting polymers (ICPs) nanoparticles is an option to improve the processability and conductivity of ICPs. In this thesis, the synthesis and application of ICPs nanoparticles has been demonstrated. Various polymerisation methods, such as emulsion polymerisation, use of steric stabiliser and synthesis in ionic liquid (IL) media, have been used to synthesise polymer nanoparticles. These synthesis methods render the ICPs nanoparticles stable as dispersions which are more processable and contain peculiar and fascinating properties superior to their bulk counterparts. These nanoparticles are further applied as mediators for biosensors. They have been fabricated into sensors using electrodeposition, evaporative casting, or ink-jet printing methods. Electrodeposition method results in formation of ultra thin nanostructured polymeric films that enhance sensor performance. Evaporative casting method is an easy one-step method, but precision is hard to achieve and dense films with rough morphology are formed. Ink-jet printing can be used to produce precise and accurate patterns and also this approach is amenable to mass production.

Polyaniline (PANI) nanoparticles; *nano*PANI-dodecylbenzene sulphonic acid (DBSA) have been synthesised using emulsion polymerisation (Chapter 3). The *nano*PANI-DBSA obtained has a conductivity of 34±7 S/cm with particle size in the range of 10±2 nm. The *nano*PANI-DBSA has been used as a mediator layer in biosensor applications as demonstrated in Chapter 7. These nanoparticles were fabricated onto the conductive electrode using an electrodeposition method with subsequent immobilisation of the enzyme horseradish peroxidase (HRP). Sensor performance was examined using

amperometric method and HRP/hydrogen peroxide (H₂O₂) configuration as a model system. The nanodomain of the *nano*PANI-DBSA particles contributed to highly ordered nanostructure patterning on the electrode surface. This uniform surface showed improved enzyme deposition characteristics, a lower background signal and better sensor performance at a lower HRP loading when compared to the sensors fabricated from electropolymerisation of the bulk monomer.

NanoPANI-DBSA particles aggregate at high concentrations; hence they are not amenable to ink-jet printing. sPANI-DBSA was prepared from centrifugation of the nanoPANI-DBSA dispersions and used as a material for ink-jet printing. HRP was premixed with the sPANI-DBSA nanodispersions before fabrication onto ITO-coated mylar using ink-jet printing. The print quality from the sPANI-DBSA nanodispersions was inconsistent and the catalytic signal of this biosensor was very low. These resulted in no further ink-jet printing work for this material.

The PANI-DBSA-rapid mixing (RM) nanodispersions were synthesised using a RM method. These dispersions contained nanometre size PANI particles dispersed in aqueous media. These nanoparticles have been successfully printed using ink-jet printing as outline in Chapter 9. This work has demonstrated the ink-jet printability of conducting polymer nanoparticles and their use as working electrodes for biosensors. The sensor response from these ink-jet printed PANI-DBSA-RM was higher than the sensor response from evaporative casting of poly(2-methoxyaniline-5-sulphonic acid) (PMAS) in Chapter 8.

The addition of functional group into PANI nanoparticles was also investigated. Carbolan Blue (CB) dye was incorporated into the PANI backbone using emulsion polymerisation method as demonstrated in Chapter 4. The dye was proved to have strong interaction with PANI backbone using Raman spectroscopy and centrifugation test. The

distinct solution colour after a reduction process could lead the PANI-DBSA-CB to be a potential candidate of the material for electrochromic devices.

Synthesis of polypyrrole (PPy) nanoparticles is demonstrated in Chapter 5. Poly(vinyl alcohol) (PVA) was used as the steric stabiliser to produce PPy-DS-PVA nanoparticles. These nanoparticles were well dispersed in water with particle size in the order of 52±5 nm. Aggregation was obvious in concentrated solutions and leaded to poor ink-jet printed quality of the PPy-DS-PVA nanoparticles.

The water soluble polymer, PMAS, was also used to fabricate biosensors using evaporative casting method in Chapter 8 and ink-jet printing in Chapter 9. In chapter 8, its solubility enabled PMAS to pre-mix with the HRP enzyme prior to complexing with the polycations poly(L-lysine) hydrochloride (PLL) and subsequently casting onto ITO coated mylar substrate. This biosensor format has proven ability to easily fabricate the conducting polymer nanoparticles by one-step evaporative casting. The optimised sensors exhibited good sensor response, high selectivity and very good long-term stability. The ink-jet printed films from PMAS and PLL solutions (Chapter 9) showed better electroactivity compared to the evaporative cast films which could lead to better sensor performance. However, the problem of PLL blocking the print head resulted in the discontinuation of its use.

The polyterthiophene (PTTh) aqueous dispersed nanoparticles were also successfully synthesised in the presence of surfactant (DBSA) and in ionic liquid; 1-ethyl-3-methylimidazolium bis(trifluoromethane-sulfonyl)amide (emiTFSA) as demonstrated in Chapter 6. The dispersion of PTTh-DBSA nanoparticles has shown poor colloidal stability and poor electroactivity. Although the PTTh nanoparticles synthesised in emiTFSA needed

2-3 minutes sonication to be dispersed in water, they have shown good electrochemistry and being test in another study in our laboratories for its use in photovoltaic devices.

These processable ICPs nanoparticles are promising materials for biosensor applications, electrochromic devices and solar cells. Assembly of these nanoparticles on to conductive substrates leads to highly ordered nanostructured ICPs on the surface and improves the biosensor performances. Also these nanoparticles prove their ability to be processable in mass production scale.