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# Surface reflectance properties of Antarctic moss and their relationship to plant species, pigment composition and photosynthetic function

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## Abstract

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## Keywords

*Bryum pseudotriquetrum*, *Ceratodon purpureus*, chlorophyll, chlorophyll fluorescence, *Grimmia antarctici*, Photochemical Reflectance Index, xanthophyll cycle pigments

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**Surface reflectance properties of Antarctic moss and their relationship to  
plant species, pigment composition and photosynthetic function**

5

RUNNING TITLE: Surface reflectance of moss

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## Abstract

We investigated how surface reflectance properties and pigment concentrations of Antarctic moss varied over species, sites, microtopography, and with water content. We found that species had significantly different surface reflectance properties, particularly in the region of the red edge (approximately 700 nm), but this did not correlate strongly with pigment concentrations. Surface reflectance of moss also varied in the visible region and in the characteristics of the red edge over different sites. Reflectance parameters, such as the Photochemical Reflectance Index (PRI) and Cold Hard Band (CBH) were useful discriminators of site, microtopographic position and water content. The PRI was correlated both with the concentrations of active xanthophyll-cycle pigments and the photosynthetic light use efficiency,  $F_v/F_m$ , measured using chlorophyll fluorescence. Water content of moss strongly influenced the amplitude and position of the red-edge as well as the PRI, and may be responsible for observed differences in reflectance properties for different species and sites. All moss showed sustained high levels of photoprotective xanthophyll pigments, especially at exposed sites, indicating moss is experiencing continual high levels of photochemical stress.

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## Introduction

Solar radiation is essential for photosynthesis and growth of plants. The surface reflectance characteristics of leaves is key in determining both the quantity and quality of solar radiation that is incident on leaf internal organs (Vogelmann 1993). Although solar radiation in the visible region drives photosynthesis, absorptance of high levels of visible radiation and radiation at other wavelengths can be damaging. At shorter wavelengths, absorbed UV-B (280-320 nm) radiation can cause lesions to nucleic acid and proteins. Excess levels of visible radiation (400-750 nm) can cause photoinhibition of photosynthesis (Cornic, Woo & Osmond 1982). High absorptance at longer wavelengths (>750 nm) leads to increases in temperature that can be detrimental in arid environments where it is important for plants to conserve water by limiting transpiration and to maintain leaf temperatures close to that of air temperatures (Gates *et al.* 1965; Ehleringer 1981). Variation in absorptance of leaves has been observed (Ehleringer & Björkman 1978; Vogelmann 1993) and specialized adaptation of surface reflectance properties in plants is known to facilitate photosynthetic carbon gain. For example, in desert ecosystems pubescent or waxy leaf surfaces that increase reflectance are common (Billings & Morris 1951, Ehleringer 1981) and have been demonstrated to be important in protection from photoinhibition of photosynthesis (e.g. Robinson, Lovelock & Osmond 1993). Moreover, epidermal tissues often accumulate compounds that are effective screens against UV-B radiation, thereby decreasing their reflectance in the UV region (e.g. Tevini 1993).

In this study we surveyed the natural variability of surface reflectance in moss in continental Antarctica. At our study site in Wilkes Land, eastern Antarctic, conditions are cold and usually very arid, resulting in lichens dominating the rocky, ice-free areas. The moss

community is well developed for the region, but it is restricted to melt lakes, melt streams and other locations where free water is available for at least some of the summer (November-March). In winter the moss is covered with snow and ice and is subject to disturbance from cryo-perturbation, which causes highly complex microrelief of ridges and valleys within the moss

5 turves. Compared to lower latitudes, levels of UV radiation are low in Antarctica. But, Antarctica is experiencing large increases in incident UV-B radiation due to reductions in concentrations of stratospheric ozone (Frederick & Snell 1988). Increases in UV radiation are having large effects on the primary production of the oceans (Neale, Davis & Cullen 1998), and could also be affecting terrestrial vegetation. The heat balance of moss, photosynthetic processes

10 and the impacts of enhanced UV radiation are likely to be partially dependent on moss surface reflectance characteristics and pigment composition. Thus, the first aim of this study was to test the hypothesis that moss species and moss inhabiting different environments have different surface reflectance characteristics. We measured surface reflectance at approximately one nm intervals from 200-900 nm of three dominant moss species over three sites with differing water

15 availabilities, and over differences in microtopography.

Surface reflectance properties of vegetation are often used as indicators of photosynthetic function in remote sensing of vegetation (reviewed in Field, Gamon & Penuelas 1994). Variation in reflectance has been observed among plant taxa (e.g. Gates *et al.* 1965; Penuelas *et al.* 1993; Gamon *et al.* 1995), due to differences in water and nutrient availability (e.g. Penuelas

20 *et al.* 1994; Gamon, Serrano & Surfus 1997), environmental stress (e.g. Rock, Hoshizaki & Millar 1988), and both seasonally (Running & Nemani 1988; Gamon *et al.* 1995) and diurnally (Gamon *et al.* 1990; Penuelas *et al.* 1994; Gamon *et al.* 1997). Changes in surface reflectance of intact leaves have been directly correlated with leaf chemical composition (Jacquemoud *et al.*

1996) and changes in photosynthetic processes in some higher plant taxa (Gamon *et al.* 1990; Gamon *et al.* 1997, Gilmore & Ball 2000). But it is often difficult to reconcile physiological measurements, which are often made at the scale of individual leaves and remotely sensed measurements, which are often made at the scale of whole canopies (Williams 1991; Field *et al.* 5 1994). Some of the difficulty arises because canopies are complex. Canopies are composed of layers of leaves of different species that can have a range of properties. Leaves within canopies can have varying distributions, orientations, morphologies, internal structures, pigment compositions, and epidermal characteristics, all properties that could influence the surface reflectance of the canopy (Rock *et al.* 1988; Field *et al.* 1994; Vose *et al.* 1995). At the leaf level, 10 mosses are very simple consisting of a few cell layers. But at a higher level of organization, moss turves, although small in comparison to canopies of trees, are relatively complex, composed of many leafy gametophytes packed together to form what can be thought of as a ‘micro-canopy’. In mosses, measurement of photosynthetic processes and surface reflectance can be made at the same scale. Thus, the second aim of our study was to test the hypothesis that 15 spectral properties of moss are directly related to photosynthetic processes and pigment composition. We wished to understand whether non-destructive hyperspectral measurements could potentially be used to map moss species distributions and physiological activity both spatially and temporally. Here we describe tests of correlations among surface reflectance properties and commonly used reflectance indices and pigment concentrations, water contents 20 and photosynthetic function of Antarctic moss.

## Materials and methods

Moss was obtained from three sites around the Australian base Casey, Wilkes Land, continental Antarctica (66°17' S, 110°32' E). Two sites were directly adjacent to the base (Red Shed and Science), while the other site was 20 km west at Robinson Ridge. Water availability varies across the sites. The Red Shed site is directly adjacent to a large summer melt lake, the Robinson Ridge site is adjacent to a melt stream that flows intermittently over the summer months, while at the Science site water is only available when overlying snow melts early in the summer. Temperature also varies over the sites. Robinson Ridge is cooler, and more exposed, than the Red Shed and Science sites (Melick, Hovendon & Seppelt 1997). In the early 1990s the Science site was exposed to contamination due to deposition of cement dust during construction (Adamson, Adamson & Seppelt 1994).

At each site representative samples, approximately 2 cm<sup>2</sup> were cut from the turves over 3 summer seasons (October to February) from 1996 to 1999. The three most common species, *Grimmia antarctici* Card., *Ceratodon purpureus* (Hedw.) Brid., and *Bryum pseudotriquetrum* (Hedw.) Gaertn., Meyer and Scherb. were sampled. The necessity of minimizing impacts of the study, the extent of the moss beds (smaller at Science) and the natural distribution of the three moss species resulted in different representations of each species at each site (Table 1). We tested for differences between moss species and the influence of site on pigment and surface reflectance parameters using samples from all sites. We used a subset of the Robinson Ridge and Red Shed *Grimmia antarctici* samples to test for effects of microtopography.

Additionally we tested the influence of moss water content on surface reflectance. We used 4 samples of *G. antarctici* from microtopographic ridges at Robinson Ridge. We measured

the surface reflectance of the samples when they were dry, and after they had been rehydrated with a fine mist of water for 5 minutes. Excess water was blotted from the moss before measurements. Moss re-hydrate rapidly, regaining full physiological activity in one to two minutes (Robinson et al. 2000).

5

### *Measuring surface reflectance*

We used an integrating sphere fitted to a scanning spectrophotometer (GBC UV-Vis 918, GBC Australia) to measure spectral reflectance of moss between 200 – 900 nm at approximately 10 1 nm intervals on 2 cm<sup>2</sup> areas of moss canopy. The moss turf samples were large enough to completely fill the sample aperture of the integrating sphere. To maintain moss canopy architecture (gametophyte arrangement) the samples were held in place by within a 1cm deep highly reflective sample holder. For some samples this required shaving tissue from the basal portion of the sample using a razor blade until the turf was 1 cm thick. The depth and packing of 15 the samples resulted in transmittance of through the sample being negligible. Reflectance measured therefore represents the moss canopy with green photosynthetic tissue of between 2-5 mm depth and 8-5 mm of non-photosynthetic tissue below. From the entire reflectance spectra, using subsets of the total 87 spectra (Table 1), we present mean reflectance at key wavelengths and also calculate reported indices used in remote sensing vegetation (Table 2). The Cold Hard 20 Band (CHB, Gilmore & Ball 2000) has been found to correlate with the formation of a chlorophyll-protein complex in leaves that protects against freezing damage. The Photosynthetic Reflectance Index (PRI, Gamon *et al.* 1990, 1997) was developed to reflect changes in concentrations of the xanthophyll cycle pigments that are formed when plants are stressed. The amplitude of the reflectance change at the red-edge ( $r_{RE}$ ) and the position of the red-edge ( $\lambda_{RE}$ )

were calculated from the first derivative of the spectra (Horler, Dockray & Barber 1983). Other reflectance indices used by a range of researchers were also calculated (e.g. Vogelmann, Rock & Moss 1993; Gitelson & Merzylak 1997; Lichtenthaler, Gitelson & Lang 1996; Carter 1991, 1993).

5

*Pigment, water and photosynthetic characteristics*

Prior to measurement of the reflectance spectra, a subset of the *G. antarctici* samples were dark adapted for 20 minutes after which the chlorophyll fluorescence parameter  $F_v/F_m$  was measured using a PAM 2000 (H. Walz, Effeltrich, Germany). After the reflectance spectra were measured, the photosynthetically active apices of the moss were removed using a razor blade and the tissue was frozen in liquid nitrogen. Samples were returned to Australia in liquid nitrogen and then stored at  $-80\text{ }^\circ\text{C}$  in a freezer prior to pigment analysis. For chlorophyll and carotenoid determination, samples (50-100 mg fresh weight) were ground with liquid nitrogen and sand in a mortar and pestle and then extracted in (1.5 ml) 100% acetone. The samples were transferred to a micro-centrifuge tube containing 1 mg sodium bicarbonate and kept on ice in the dark for 20 minutes. After centrifugation (13,000 rpm for 5 minutes) the pellet was re-extracted in 0.5 ml 80% acetone using a polypropylene tissue grinder (Crown Scientific, Australia). After a further 20 minutes on ice, the sample was centrifuged as above and the supernatants from each extraction combined and made up to 3 ml with 100% acetone. Chlorophylls and carotenoids were then quantified by high-pressure liquid chromatography (HPLC) using a method adapted from Gilmore & Yamamoto (1991). The samples (30-70  $\mu\text{l}$ ) were injected into a Shimadzu HPLC system (Shimadzu, Australia) at a flow rate of  $2\text{ ml min}^{-1}$ . Solvent A

(acetonitrile:methanol:Tris HCL buffer 0.1 M, pH 8.0; 79:8:3) ran isocratically from 0 to 4 min followed by a 2.5 min linear gradient to 100% Solvent B (methanol:hexane; 4:1) which then ran isocratically from 6.5 -15 min. Flow rate was decreased from 2 to 1.5 ml min<sup>-1</sup> between 6-12 min and then ran at 1.5 ml min<sup>-1</sup> until 15 min, to maintain stable pressure. The column was re-  
5 equilibrated with solvent A between samples. Pigments were separated on an Spherisorb ODS1 column (Alltech, Australia) and quantified by integration of peak areas, detected at 440 nm using a photodiode array detector (Model SPD-M10AVP, Shimadzu, Australia), relative to pure chlorophyll (Sigma, Australia) and carotenoid (Extrasynthase, France and VKI, Denmark) standards.

10 Anthocyanin concentrations were determined using the differential pH method of Fuleki & Francis (1968) and Francis (1982). Moss tissue (0.1-0.2 g fresh weight) was ground with liquid nitrogen and sand in a mortar and pestle and then extracted in (1.5 ml) of 1% HCl in methanol. Samples were centrifuged at 13,000 rpm for 6 min and absorbance of the supernatant measured at 526nm. 100 µl of sodium acetate buffer (pH 5.0) was then added to 1 ml of the  
15 supernatant and after 3 min this sample was centrifuged for 10 min at 13,000 rpm before a second measurement of absorbance at 526 nm. Anthocyanin concentration was calculated from the difference in these two absorbance measures.

Water content of samples was measured after oven drying at 80 °C to stable weight and is expressed as gH<sub>2</sub>O gdryweight<sup>-1</sup> as described in Robinson *et al.* (2000). Concentration of UV-B  
20 absorbing pigments was determined from 15 to 60 mg of dry moss tips. Dry tissue was weighed, ground in liquid nitrogen with acid-washed sand and then extracted in 1.5 ml of acidified methanol (methanol-H<sub>2</sub>O-HCl; 79:20:1). The sample was transferred to a microfuge tube and allowed to stand on ice in the dark, with vortexing every 20 min. After 1 hour the tube was

centrifuged (13,000 rpm, 4 min) and the supernatant removed. The final volume of supernatant was made up to 1.5 ml with acidified methanol. Absorbance across the UV-B region (280-320nm) was scanned spectrophotometrically. Values are expressed as mean absorbance for this wavelength range.

5

### *Data analysis*

Correlations among surface reflectance indices, pigments, chlorophyll fluorescence parameter  $F_v/F_m$  and water content were tested using the non-parametric Spearman Rank Correlation (*Rho*) using the statistical computing package Data Desk 6.1 (Data Descriptions 6.1, Ithaca NY). *Rho* detects consistently increasing or decreasing trends but does not assume variables are linearly related. Linear regression was also used to test for significant relationships between moss chlorophyll concentrations and reflectance indices, and among other variables. Tests of the influence of site, species and microtopography on pigments and reflectance parameters were performed using analysis of variance (ANOVA). For tests of sites and species ANOVAs were unbalanced (unequal sample sizes), therefore post-hoc tests of differences between individual means were adjusted for the unequal sample sizes using the Bonferroni test. The adequacy of the ANOVA models was assessed by inspecting residual plots. Paired t-tests were used to assess the influence of hydration on reflectance parameters.

20

## Results

### *Differences between species in reflectance and pigments*

5 Reflectance spectra of mosses were similar in shape to those observed for intact leaves of angiosperms with the major features of the spectra those indicative of chlorophyll (Fig 1). Examination of the 1<sup>st</sup> derivative of the spectra (Fig 1B) showed typical peaks at the green (approximately 525 nm) and red edge (approximately 700 nm). Moss species did not significantly differ in their UV reflectance (R320), but they differed significantly at 526, 550 and 10 850 nm, in both  $R_{RE}$  and  $R_{RE'}$  and tended to have different CHB (Table 3). In all cases, *B. pseudotriquetrum* had greater reflectance values.

Pigment concentrations also varied among species (Table 3). *Bryum pseudotriquetrum* had higher levels of UV-absorbing pigments but lower carotenoid levels (total carotenoid and xanthophyll concentrations) compared to *C. purpureus* and *G. antarctici*. *Ceratodon purpureus* 15 had lower total chlorophyll concentrations than *G. antarctici* or *B. pseudotriquetrum* but higher levels of anthocyanins.

Many of the moss pigments concentrations tended to co-vary with each other. For example, total chlorophyll concentration was highly correlated with the total carotenoid content ( $Rho = 0.91$ ). Additionally, concentrations of all the carotenoids tended to be highly correlated 20 (e.g. correlation among lutein and the xanthophyll cycle pigments  $Rho = 0.726$ ). Anthocyanins, measured as absorbance of extracts at 526 nm and expressed on a fresh wt basis, were also correlated with total chlorophyll ( $Rho = 0.625$ ). The proportion of xanthophyll-cycle pigments in

the de-epoxidated photoprotective forms, zeaxanthin and antheraxanthin, was negatively correlated with total chlorophyll ( $Rho = -0.482$ ) and positively correlated with the total pool size of the xanthophyll-cycle pigments ( $Rho = 0.588$ ).

## 5 *Microtopography*

The PRI was strongly influenced by whether moss was growing on a ridge or valley, but no other reflectance parameter was significantly affected by microtopographic position (Table 4).

Samples from ridges had lower values of PRI (more negative) than those in the valleys. Total chlorophyll concentrations were approximately half on the ridges compared to the valleys, while the concentration of xanthophyll-cycle pigments per chlorophyll was 50% higher. There was also a significantly higher proportion of the xanthophyll-cycle present in the photoprotective forms on ridges.

## 15 *Sites*

Over the three sites we observed significant difference in many of the reflectance parameters, but there were no significant effects of site on R320, PRI, NDVI or NPCI (Table 5). The Science site had higher reflectance at 526 and 550 nm, but lower reflectance at 850 nm compared to moss growing at the Red Shed and Robinson Ridge sites. Moss at the Red Shed site exhibited greater  $R_{RE}$  and  $R_{RE}$  values than did moss at the Robinson Ridge and Science sites. The CHB was also higher at Robinson Ridge than at the other two sites. Moss at the Red Shed site had lower EGFR and EGFN than moss at Robinson Ridge and Science.

Differences in reflectance indices corresponded to changes in pigment contents over the sites. Total chlorophyll and anthocyanin concentrations were not significantly different among sites, although mosses tended to have higher concentrations of these pigments at Robinson Ridge than at Science and Red Shed sites (Table 5). Xanthophyll pigment concentrations, total  
 5 carotenoids and concentrations of UV absorbing pigments were all greater at Robinson Ridge than at the other two sites.

*Relationships among reflectance, plant pigments, water content and photosynthesis*

10 Overall, correlation between surface reflectance indices and plant pigment contents was low. Concentration of total chlorophyll was correlated significantly, but not strongly with many of the key reflectance wavelengths and indices (Table 6). The strongest linear regression was between total chlorophyll and reflectance indices calculated with reflectance measured at 850 and 680 nm (either R850, R850-680, and NDVI) and with  $r_{RE}$ . There was no significant relationship  
 15 between total chlorophyll and EGFR, EGFN or NPCI.

The surface reflectance characteristic of moss that correlated most strongly with the concentration of extracted anthocyanin pigments measured at 526 was  $r_{RE}$  ( $Rho = 0.426$ ). The PRI, a tool developed to remotely assess plant stress (Gamon *et al.* 1997) was negatively correlated with both the total pool size of the xanthophyll-cycle pigments ( $Rho = -0.402$ ) and  
 20 also the proportion as zeaxanthin and antheraxanthin (Fig 3A,  $Rho = -0.535$ ). The PRI was positively correlated with the chlorophyll fluorescence parameter,  $F_v/F_m$  (Fig 3B,  $Rho = 0.626$ ). Declines in  $F_v/F_m$  indicate declines in the efficiency of photosystem II.

To assess the influence of water content on reflectance parameters we rehydrated dry samples of *G. antarctici* in the laboratory. Re-hydration resulted in a significant increase in  $R_{RE}$  accompanied by a shift in  $R_{RE}$  to shorter wavelengths (Table 7). Re-hydration also resulted in a significant increase in the PRI and a decrease in the CHB (Table 7).

5

## Discussion

### *Species*

10 We found that moss species, moss at different sites, and moss from different microtopographic positions varied significantly in their surface reflectance properties, but different reflectance parameters were sensitive to different ecological or physiological factors (Table 8). The parameters most sensitive to species and site factors were reflectance in the visible region (R526 and R550), and the features of the red-edge ( $R_{RE}$  and  $R_{RE}$ ). Among moss species, *B.*

15 *pseudotriquetrum*, had higher values of reflectance indices than other species (Fig. 1, Table 3). This was not simply associated with differences in chlorophyll concentrations, which were higher than those of *C. purpureus* but similar to that measured in *G. antarctici*. *Bryum pseudotriquetrum* is morphologically distinct from the other two species, having much larger gametophytes with larger leaves that are densely packed into the turf (S.A. Robinson,

20 unpublished data). Gametophyte size is similar in the other two species, but gametophytes of *C. purpureus* are also densely packed, whereas those of *G. antarctici* plants are more loosely arranged within the turves. Reflectance parameters are likely to be influenced by gametophyte morphology and packing independently of chlorophyll concentrations. Reflectance from

canopies is sensitive to the leaf area index (LAI; Booches *et al.* 1990; Penuelas *et al.* 1994; Serrano, Gamon & Penuelas 2000) and to other features of canopy geometry e.g. leaf thickness, surface quality and orientation (Rock *et al.* 1988). Thus, differences observed in reflectance characteristics among moss species could be more strongly influenced by differences in plant morphology (e.g. gametophyte morphology, density and thickness of photosynthetic tissue) than by physiological factors. It should be noted that *B. pseudotriquetrum* is the species that shows the most ‘angiosperm leaf like’ reflectance. Unfortunately, since moss surface reflectance has not been well studied we are unable to say if *B. pseudotriquetrum* or the other species are most typical of moss plants.

10           In previous work we have found that these three moss species desiccate at different rates and can maintain photosynthetic function at different levels of desiccation (Robinson *et al.* 2000). Given the sensitivity of the red-edge features to water content (Table 7), differences among reflectance of moss species and sites could also be due to variation in water contents.

          Pigment composition also varied between species with *B. pseudotriquetrum*, having  
15 higher levels of specific UV –screening pigments whilst *C. purpureus* and *G. antarctici* showed higher levels of pigments generally associated with photoprotection from excess visible light. In the latter species the ratio of xanthophyll cycle pigments/chlorophyll was high and characteristic of plants exposed to much higher levels of solar radiation. Given that samples for pigment  
20 analysis were not frozen until several hours after collection, the sustained de-epoxidation status of the xanthophyll cycle pool is also indicative of relatively high photochemical stress in these plants. Differences in relative pigment concentrations between the three species also suggests that they may be differentially vulnerable to increasing UV-B exposure as a result of ozone depletion.

### *Microtopography*

Over different microtopographic positions *G. antarctici* samples on ridges had lower PRI than  
5 samples from valleys, but no differences were observed in any other reflectance parameters,  
despite large differences in chlorophyll concentrations between moss on ridges and valleys. In  
other studies higher chlorophyll concentrations and proportion of green tissue are associated with  
shifts in  $\tau_{RE}$  to longer wavelengths (Curran *et al.* 1992; Vogelmann *et al.* 1993; Serrano *et al.*  
2000). The lack of difference in the features of the red-edge, or other reflectance indices despite  
10 large differences in chlorophyll concentrations over ridges and valleys suggest that  $\tau_{RE}$  and  $\rho_{RE}$   
are largely independent of chlorophyll concentrations over the range found in this study. In both  
ridges and valleys moss turves form closed micro-canopies that likely have high 'leaf' area  
indices (LAI). In higher plants, canopies with high LAI are often not distinguishable using  
reflectance parameters (Booches *et al.* 1990; Penuelas *et al.* 1994; Serrano, Gamon & Penuelas  
15 2000). Thus, the dense packing of gametophytes in moss may contribute to the insensitivity of  
reflectance parameters to differences in microtopography.

Lower PRI on ridges than valleys was associated with higher concentrations of  
xanthophyll-cycle pigments and greater de-epoxidation of xanthophyll cycle pigments;  
characteristics of plants experiencing conditions that limit photosynthetic carbon gain and result  
20 in photoinhibition (Osmond 1994; Björkmann & Demmig-Adams 1995; Lovelock, Osmond &  
Seppelt 1995a; Lovelock *et al.* 1995b). Moss on ridges is exposed to much higher levels of solar  
radiation than moss shaded in microtopographic valleys. Moss on ridges also experience lower  
and more variable water availability. Higher light levels in combination with lower water

contents are likely to lead to photoinhibition, de-epoxidation of xanthophyll cycle pigments and reduced PRI.

### *Sites*

5

Those reflectance indices sensitive to site but not species were EGFN, EGFR and CHB. In our assessment of reflectance parameters over sites we expected that sites with lower water availability (Robinson Ridge and Science) would have differences in reflectance parameters and pigment concentrations that were indicative of greater exposure to environmental stress. Our prediction was largely supported with EGFN and EGFR being higher at the drier Science and Robinson Ridge sites, than at the Red Shed site (Table 6). Differences in EGFN and EGFR are products of the differences in the red edge over sites, where the amplitude of the red edge is steeper and at longer wavelengths at the drier sites. The CHB was greater at the cooler western Robinson Ridge site than at Science and Red Shed, which corresponded to the lower average temperatures recorded at Robinson Ridge (Melick *et al.* 1997).

15

Total chlorophyll concentrations were greater at Robinson Ridge than at the other sites. Because we did not observe differences in  $_{RE}$  and  $_{RE}$  due to the microtopographic position of moss, where chlorophyll concentrations were vastly different, we conclude that changes in  $_{RE}$  and  $_{RE}$  over sites is due to some other factors than differences in chlorophyll concentrations. Other important factors could be differences in moss species composition over the sites, differences in water content, and/or gametophyte morphology. It is also possible that the higher reflectance at the Science site is also due to the increased presence of fine dust particles on the moss surface as a result of local human activity.

20

*Relationships among reflectance, plant pigments, water content and photosynthesis*

Despite sensitivity of surface reflectance characteristics to species, site and microtopography, surface reflectance indices were not strongly correlated with pigment concentrations, with the exception of the correlation between PRI and xanthophyll-cycle pigments. The PRI also correlated with the chlorophyll fluorescence parameter  $F_v/F_m$ , a measure of PSII function, as has also been shown by Gamon *et al.* (1997) and others (e.g. Méthy 2000). The NDVI, an index commonly used in broad band remote sensing, was shown to be insensitive to the range of factors assessed in this study (Table 8) as has also been found by others (e.g. Buschmann & Nagel 1993).

Water content of moss appears to be a key parameter influencing  $R_{RE}$  and  $R_{RE}$  and PRI. The red edge has been shown to shift to longer wavelengths when bean leaves are infiltrated by water (Knipling 1970). More commonly observed is a 'blue shift' in leaves and plant that are subjected to water stress (Horler *et al.* 1983; Booches *et al.* 1990; Carter 1991, 1993; Penuelas & Inoue 1999), although in some cases measurements are complicated by a reduction in chlorophyll in the water stressed tissues (Maracci *et al.* 1991). Desiccation tolerant mosses show much greater variation in water content than angiosperm leaves (i.e. these species tolerate a 12- 28 fold difference in water content in live tissues, Robinson *et al.* 2000). Thus, a shift toward the red as water content increases in moss may reflect processes that are particular to the physiology of desiccation resistant plants. These processes may be more similar to those involved in greening of angiosperm leaves (e.g. Gates *et al.* 1965; Curren *et al.* 1991; Vogelmann *et al.* 1993) than those as a result of acute environmental stress.

The sensitivity of the surface reflectance indices to species and site and some physiological parameters suggests that mapping of moss communities might be possible using hyperspectral sensors. From these results it seems likely that high spectral and spatial resolution sensor such as the Compact Airborne Spectrographic Imager (CASI), programmed to detect narrow wavebands in the visible and NIR, could potentially differentiate between species and detect changes in physiological activity at small spatial scales (1 m<sup>2</sup> or less, depending on the deployment height of the sensor). These methods have already proven useful in the mapping of aquatic and littoral communities, where the use of visible wavebands was essential (Zacharias *et al.*, 1992; Bajjouk, Guillaumont & Poplus 1996, Fyfe and Dekker 2001) and in assessment of photosynthetic activity in terrestrial plant communities (e.g. Zarco-Tejada *et al.* 2000). Hyperspectral remote sensing could provide an effective and nondestructive method of mapping the moss communities that occur in remote Antarctic and other high latitude locations, and to monitor impacts of climate change on the spatial extent and composition of these communities that are highly sensitive to climatic trends (Doran *et al.* 2001).

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### *Conclusions*

In Antarctic moss surface reflectance characteristics vary among species and are significantly influenced by the environmental conditions under which plants grow. Differences in reflectance characteristics across sites and species were not clearly linked to changes in pigment concentrations but maybe more strongly linked to differences in water contents and morphology of gametophytes and turves. The PRI was the surface reflectance index most sensitive to variation in pigment compositions and photosynthetic function. Hyperspectral imaging of plant

20

communities? may be a useful tool in assessing the impacts of global climate change in Antarctica. Moss may also be a useful 'system' to test the applicability of new parameters for remotely sensing the physiological state of vegetation.

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## References

- Adamson E., Adamson H. & Seppelt R.D. (1994) Cement dust contamination of *Ceratodon*  
15 *purpureus* at Casey, East Antarctica: damage and capacity for recovery. *Journal of Bryology* **18**, 127-137.
- Bajjouk T., Guillaumont B. & Poplus J. (1996) Application of airborne imaging spectrometry  
system data to intertidal seaweed classification and mapping. *Hydrobiologia* **326**, 463-  
471.
- 20 Billings, W.D. & Morris, R.J. (1951) Reflection of visible and infrared radiation from leaves of  
different ecological groups. *American Journal of Botany* **38**, 327-331.
- Björkman O. & Demmig-Adams B. (1995) Regulation of photosynthetic light energy capture,  
conversion and dissipation in leaves of higher plants. In: *Ecophysiology of*

Photosynthesis (eds. E. D. Schulze & M. M. Caldwell), pp 17-47, Springer-Verlag, Berlin.

Booches F., Kupfer G., Dockter K. & Kühbauch W. (1990) Shape of the red edge as a vitality indicator for plants. *International Journal of Remote Sensing* **11**, 1741-1753.

5 Buschmann C. & Nagel E. (1993) In vivo spectroscopy and internal optics of leaves as a basis for remote sensing of vegetation. *International Journal of Remote Sensing*, **14**, 711-722.

Carter G.A. (1991) Primary and secondary effects of water content on the spectral reflectance of leaves. *American Journal of Botany* **78**, 916-924.

Carter G.A. (1993) Responses of leaf spectral reflectance to plant stress. *American Journal of*  
10 *Botany* **80**, 239-243.

Cornic, G., Woo, K.C. & Osmond C.B. (1982) Photoinhibition of CO<sub>2</sub> (carbon dioxide)-dependent O<sub>2</sub> (oxygen) evolution by intact chloroplasts isolated from spinach leaves (*Spinacia oleracea*). *Plant Physiology* **70**, 1310-1315.

Curran P.J., Dungan J.L. & Gholz H.L. (1992) Seasonal LAI in slash pine estimated with  
15 Landsat TM. *Remote Sensing of Environment* **39**, 3-13.

Doran P.T., Priscu J.C., Lyons W.B., Walsh J.E., Fountain A.G., McKnight D.M., Moorhead D.L., Virginia R.A., Wall D.H., Clow G.D., Fritsen C.H., McKay C.P. & Parsons A.N. (2002) Antarctic climate cooling and terrestrial ecosystem response. *Nature* **415**: 517-520.

20 Ehleringer J. (1981) Leaf absorptances of Mohave and Sonoran Desert plants. *Oecologia* **49**, 366-370.

Ehleringer J. & Björkman O. (1978 ) Pubescence and leaf spectral characteristics in a desert shrub *Encelia farinosa*. *Oecologia* **36** 151-162.

- Field C.B., Gamon J.A. & Penuelas, J. (1994) Remote sensing of terrestrial photosynthesis. In: Ecological studies: analysis and synthesis 100: (eds ???) pp 511-527. Springer-Verlag, New York.
- Francis F.J. (1982) Analysis of anthocyanins. In: Anthocyanins as food colours (ed. P. Markakis), pp 181-207. Academic Press, New York.
- 5 Frederick J.E. & Snell H.E. (1988) Ultraviolet radiation levels during the Antarctic spring *Science* **241**, 438-9.
- Fuleki T. & Francis F.J. (1968) Quantitative methods for anthocyanins 2. Determination of total anthocyanin and degradation index for Cranberry juice. *Journal of Food Science* **33**, 78-10 83.
- Fyfe S.K. & Dekker, A.G. (2001) Seagrass species are they spectrally distinct? In: IEEE 2001 International Geoscience and Remote Sensing Symposium, 9-13 July, 2001, Sydney, Australia, Vol. 6, pp.2740-2742..
- Gamon J.A., Field, C.B., Bilger W., Björkman O., Fredeen A.L. & Penuelas J. (1990) Remote 15 sensing of the xanthophyll-cycle and chlorophyll fluorescence in sunflower leaves and canopies *Oecologia* **85**, 1-7.
- Gamon J.A., Field, C.B., Goulden, M.J., Griffen K.L., Harley A.E., Joel G, Penuelas J. & Valentini R. (1995) Relationship between NDVI, canopy structure, and photosynthesis in three Californian vegetation types. *Ecological Applications* **5**, 28-41.
- 20 Gamon J.A., Serrano L. & Surfus J.S. (1997) The photochemical reflectance index: an optical indicator of photosynthetic radiation use efficiency across species and functional types, and nutrient levels. *Oecologia* **112**, 492-501.

- Gates D.M., Keegan H.J., Schleter J.C. & Weidner V.R. (1965) Spectral properties of plants. *Applied Optics* **4**, 11-20.
- Gilmore A.M. & Ball M.C. (2000) Protection and storage of chlorophyll in overwintering evergreens. *Proceedings of the National Academy of Science*. **97**, 11098-11101.
- 5
- Gilmore A.M. & Yamamoto H.Y. (1991). Resolution of lutein and zeaxanthin using a nonendcapped, lightly carbon-loaded C<sub>18</sub> high-performance liquid chromatography column. *Journal of Chromatography* **543**, 137-145.
- Gitelson A.A. & Merzlyak M.N. (1997) Remote estimation of chlorophyll content in higher plant leaves. *International Journal of Remote Sensing* **18**, 2691-2697.
- 10
- Jacquemoud S., Ustin S.L., Verdebout J., Schmuck G., Andreoli G., & Hosgood B. (1996) Estimating leaf biochemistry using PROSPECT leaf optical properties model. *Remote Sensing of Environment* **56**, 194-202.
- Horler D.N.H., Dockray M. & Barber J. (1983) The red edge of leaf reflectance *International Journal of Remote Sensing* **4**, 273-583.
- 15
- Knipling E.B. (1970) Physical and Physiological basis for the reflectance of visible and near-infrared radiation from vegetation. *Remote Sensing of Environment* **1**, 155-9.
- Lichtenthaler H.K., Gitelson A. & Lang M. (1996) Non-destructive determination of chlorophyll content of leaves of a green and an aurea mutant of tobacco by reflectance measurements. *Journal of Plant Physiology* **148**, 483-493.
- 20
- Lovelock C.E., Osmond C.B., & Seppelt R.D. (1995a) Photoinhibition in the Antarctic moss *Grimmia antarctici* Card. when exposed to cycles of freezing and thawing. *Plant, Cell and Environment* **18**, 1395-1402.

- Lovelock C.E., Jackson A.E., Melick D.R., & Seppelt R.D. (1995b) Reversible photoinhibition in Antarctic moss during freezing and thawing. *Plant Physiology* **109**, 955-961.
- Lichtenthaler H.K., Gitelson A. & Lang M. (1996) Non-destructive determination of chlorophyll content of leaves of a green and an aurea mutant of tobacco by reflectance measurements. *Journal of Plant Physiology* **148**, 483-493.
- Maracci G., Schmuck G., Hosgoog B. & Andreoli G. (1991) Interpretation of reflectance spectra by plant physiological parameters. In: IGARSS Remote sensing global monitoring for Earth management. (eds. ??) pp 2303-2306 Pub, city.
- Melick DR, Hovendon M, & Seppelt R (1997) Vegetation patterns in relation to climatic and endogenous changes in Wilkes Land, continental Antarctica. *Journal of Ecology* **85**, 43-56.
- Méthy M. (2000) A two-channel hyperspectral radiometer for the assessment of photosynthetic radiation-use efficiency. *Journal of Agricultural Engineering Research* **75**, 107-110.
- Osmond C.B. (1994) What is photoinhibition? Some insights from comparison of sun and shade plants. In: Photoinhibition: molecular mechanisms to the field. (eds. N.R. Baker & J.R. Boyer), pp 1-24. Bios Scientific Publications, Oxford.
- Neale P.J. Davis R.F. & Cullen J.J. (1998) Interactive effects of ozone depletion and vertical mixing on photosynthesis of Antarctic phytoplankton. *Nature* **392**, 585-589.
- Penuelas J., Gamon J.A., Fredeen A.L., Merino J. & Field C.B. (1994) Reflectance indices associated with physiological changes in nitrogen- and water-limited sunflower leaves. *Remote Sensing of Environment* **48**, 135-146.

- Penuelas J., Gamon J.A., Griffin K.L. & Field C.B. (1993) Assessing community type, plant biomass, pigment composition, and photosynthetic efficiency of aquatic vegetation from spectral reflectance. *Remote Sensing of Environment* **46**, 110-118.
- Penuelas, J. & Inoue, Y. (1999) Reflectance indices indicative of changes in water and pigment contents of peanut and wheat leaves. *Photosynthetica* **36**, 355-360.
- Robinson S.A., Wasley J., Popp M. & Lovelock C.E. (2000) Desiccation tolerance of three moss species from continental Antarctica. *Australian Journal of Plant Physiology* **27**, 379-388.
- Robinson S.A., Lovelock C.E. & Osmond C.B. (1993) Wax as a mechanism for protection against photoinhibition: a study of *Cotyledon orbiculata*. *Botanica Acta* **106**, 307-312
- 10 Rock B.N., Hoshizaki T. & Miller J.R. (1988) Comparison of *in situ* and airborne spectral measurements of the blue shift associated with forest decline. *Remote Sensing of Environment* **24**, 109-127.
- Rouse, J.W., R.H. Haas, J.A. Schell & Deering D.W. (1973) Monitoring vegetation systems in the Great Plains with ERTS. *Proceedings 3rd ERTS Symposium*, Vol. 1, pp.48-62.
- 15 Running S.W. & Nemani R.R. (1988) Relating seasonal patterns of the AVHRR index to simulated photosynthesis and transpiration of forests in different climates. *Remote Sensing of Environment* **24**, 347-367.
- Serrano L., Gamon J.A. & Penuelas, J. (2000) Estimation of canopy photosynthetic and nonphotosynthetic components from spectral transmittance. *Ecology* **81**, 3149-3162.
- 20 Tevini M. (1993) Effects of enhanced UV-B radiation on terrestrial plants. In: UV-B radiation and ozone depletion. Effects on humans, animals, plants, microorganisms and materials (ed M. Tevini), pp 125-153. CRC Press Inc, Boca Raton.

- Vogelmann T.C. (1993) Plant tissue optics. *Annual Review of Plant Physiology and Plant Molecular Biology* **44**, 231-251.
- Vogelmann J.E., Rock B.N. & Moss D.M. (1993) Red edge spectral measurements from sugar maple leaves. *International Journal of Remote Sensing* **14**, 1563-1575.
- 5 Vose J.M., Sullivan N.H. Clinton B.D. & Boldstad P.V. (1995) Vertical leaf area distribution, light transmittance, and application of the Beer-Lambert law in four mature hardwood stands in the southern Appalachian. *Canadian Journal of Forest Research* **25**, 1036-1043.
- Williams D.L. (1991) A comparison of spectral reflectance properties at the needle, branch and canopy levels for selected conifer species. *Remote Sensing of Environment* **35**, 79-93.
- 10 Yoder B. & Waring R.H. (1994) The normalized difference vegetation index of small Douglas fir canopies with varying chlorophyll concentrations. *Remote Sensing of Environment* **49**, 81-91.
- Zacharias M., Niemann O. & Borstad G. (1992) An assessment and classification of a multispectral bandset for the remote sensing of intertidal seaweeds. *Canadian Journal of Remote Sensing* **18**, 263-274.
- 15 Zarco-Tejada P.J., Miller J.M., Mohammed G.H., Noland T.L. & Sampson P.H. (2000) Chlorophyll fluorescence effects on vegetation apparent reflectance: II Laboratory and airborne canopy-level measurements with hyperspectral data. *Remote Sensing of Environment* **74**, 596-608.

Table 1. Details of species and site for moss samples used in the study.

Species	Number of Samples per Site		
	Robinson Ridge	Red shed	Science
<i>Ceratodon purpureus</i>	4	4	5
<i>Bryum pseudotriquetrum</i>	7	3	2
<i>Grimmia antarctici</i>	24	36	2
TOTAL	35	43	9

Table 2. Calculation of reflectance indices

Parameter	Index	Calculation
Cold Hard Band <sup>1</sup>	CHB	$(R750-R710)/(R750+R710)$
Normalized difference between the first derivative in the red and green <sup>2</sup>	EGFN	$(1^{st} R530-1^{st} R710)/(1^{st} R530+1^{st} R710)$
Ratio of first derivative in the green to first derivative in the red <sup>2</sup>	EGFR	$(1^{st} R530/1^{st} R710)$
Normalized difference vegetation index <sup>3</sup>	NDVI	$(R850-R680)/(R850+R680)$
Normalized pigment chlorophyll ratio index <sup>2</sup>	NPCI	$(R680-R430)/(R680+R430)$
Photosynthetic reflectance index <sup>4</sup>	PRI	$(R531-R570)/(R531+R570)$

<sup>1</sup>(Gilmore and Ball 2000); <sup>2</sup>(Penuelas *et al.* 1994); <sup>3</sup>(Rouse *et al.* 1973); <sup>4</sup>(Gamon *et al.* 1997)

Table 3. Surface reflectance characteristics and pigment concentrations for three common moss species in Continental Antarctica, N = 11, 12, 63 respectively. NS signifies a non-significant difference at the  $P=0.10$  level).

	<i>Bryum pseudotriquetrum</i>	<i>Ceratodon purpureus</i>	<i>Grimmia antarctici</i>	F	P
<b>Reflectance</b>					
R320	0.0070 ± 0.0008	0.0071 ± 0.0010	0.0070 ± 0.0003		NS
R526	0.049 ± 0.005	0.022 ± 0.006	0.023 ± 0.001	28.40	<0.0001
R550	0.055 ± 0.007	0.034 ± 0.008	0.034 ± 0.002	5.84	0.0074
R850	0.388 ± 0.037	0.275 ± 0.020	0.311 ± 0.012	4.29	0.0170
RE	0.0101 ± 0.0001	0.0060 ± 0.0001	0.0068 ± 0.0001	8.74	0.0004
ARE	705.4 ± 1.9	700.6 ± 1.0	700.4 ± 0.5	7.33	0.0012
PRI	-0.160 ± 0.028	-0.160 ± 0.015	-0.172 ± 0.009		NS
NDVI	0.835 ± 0.047	0.800 ± 0.029	0.868 ± 0.012		NS
NPCI	0.442 ± 0.057	0.484 ± 0.043	0.398 ± 0.024		
EGFR	6.04 ± 1.58	8.82 ± 2.43	8.03 ± 0.85		NS
EGFN	0.643 ± 0.042	0.784 ± 0.072	0.701 ± 0.021		NS
CHB	0.349 ± 0.058	0.229 ± 0.021	0.261 ± 0.015	2.74	0.0708
<b>Pigments</b>					
Anthocyanins	1.2 ± 0.2	1.3 ± 0.2	1.2 ± 0.2		NS
$A_{526 \text{ diff}} \text{ g}^{-1} \text{ fwt}$					
Anthocyanins/TChl.	2.9 ± 0.3	5.7 ± 1.2	3.0 ± 0.2	7.42	0.0011
$A_{526 \text{ diff}} \text{ mol}^{-1}$					
UV absorbing pigments	266 ± 42	127 ± 22	110 ± 9	15.34	<0.0001
Mean $A_{(320-280\text{nm})} \text{ g}^{-1} \text{ dwt}$					
Total Chlorophyll	475 ± 60	241 ± 40	453 ± 44	2.50	0.0882
$\text{nmol g}^{-1} \text{ fwt}$					
Chl a:b ratio	3.2 ± 0.2	3.6 ± 0.3	3.2 ± 0.1		NS
VAZ/TChl	64 ± 5	132 ± 36	103 ± 6	3.64	0.0305
$\text{mmol mol}^{-1}$					
%(AZ/VAZ)	18.0 ± 3.9	39.4 ± 4.9	37.4 ± 2.5	6.37	0.0026
Tcar./Tchl.	443 ± 15	643 ± 96	631 ± 31	2.86	0.0629
$\text{mmol mol}^{-1}$					

Table 4. Surface reflectance properties and pigment concentrations of *Grimmia antarctici* over different microtopographic positions. Moss is from ridges or valleys at Robinson Ridge and Red Shed sites,  $N=8$ . NS signifies a non-significant difference at the  $P=0.10$  level).

	Ridge	Valley	<i>F</i>	<i>P</i>
<b>Reflectance</b>				
R320	0.0058 ± 0.0003	0.0066 ± 0.0006	NS	
R526	0.027 ± 0.003	0.0022 ± 0.001	NS	
R550	0.035 ± 0.004	0.042 ± 0.003	NS	
R850	0.362 ± 0.016	0.374 ± 0.023	NS	
RE	0.0079 ± 0.0001	0.0084 ± 0.0001	NS	
λRE	700.75 ± 1.26	700.25 ± 1.19	NS	
PRI	-0.213 ± 0.009	-0.117 ± 0.011	48.79	0.0001
NDVI	0.849 ± 0.013	0.882 ± 0.031	NS	
NPCI	0.333 ± 0.064	0.425 ± 0.069	NS	
EGFR	8.90 ± 1.77	9.33 ± 2.16	NS	
EGFN	0.761 ± 0.032	0.780 ± 0.043	NS	
CHB	0.295 ± 0.025	0.256 ± 0.036	NS	
<b>Pigments</b>				
Anthocyanins	1.0 ± 0.4	1.9 ± 0.3	NS	
$A_{526 \text{ diff}} \text{g}^{-1} \text{fw}$				
Anthocyanins/TChl.	2.7 ± 0.5	2.6 ± 0.6	NS	
$A_{526 \text{ diff}} \text{mol}^{-1}$				
Total Chlorophyll	375 ± 73	791 ± 66	17.88	0.0008
$\text{nmol g}^{-1} \text{fw}$				
Chl a:b ratio	3.3 ± 0.2	3.1 ± 0.2	NS	
VAZ/TChl	130 ± 12	81 ± 6	12.87	0.0030
$\text{mmol mol}^{-1}$				
%(AZ/VAZ)	40.9 ± 5.4	27.1 ± 3.3	4.82	0.0454
Tcar./Tchl.	761 ± 87	465 ± 18	10.98	0.0051
$\text{mmol mol}^{-1}$				

Table 5. Surface reflectance characteristics and pigment concentrations of moss over three sites in Continental Antarctica. N = 34, 43, 9 respectively. NS signifies a non-significant difference at the  $P=0.10$  level).

	Robinson Ridge	Red shed	Science	<i>F</i>	<i>P</i>
<b>Reflectance</b>					
R320	0.0068 ± 0.0004	0.0070 ± 0.0004	0.0086 ± 0.0011	NS	
R526	0.0027 ± 0.0004	0.0023 ± 0.0004	0.0038 ± 0.0011	4.94	0.0094
R550	0.035 ± 0.002	0.034 ± 0.001	0.052 ± 0.004	3.43	0.0371
R850	0.351 ± 0.017	0.289 ± 0.015	0.331 ± 0.024	4.28	0.0170
RE	0.0081 ± 0.0001	0.0063 ± 0.0001	0.0070 ± 0.0001	3.93	0.0235
ARE	703.8 ± 1.1	699.3 ± 0.2	700.1 ± 0.4	11.91	<0.0001
PRI	-0.165 ± 0.014	-0.177 ± 0.010	-0.146 ± 0.011	NS	
NDVI	0.857 ± 0.019	0.864 ± 0.015	0.808 ± 0.042	NS	
NPCI	0.454 ± 0.035	0.391 ± 0.027	0.370 ± 0.051	NS	
EGFR	10.19 ± 1.43	5.86 ± 0.58	9.30 ± 2.99	4.62	0.0127
EGFN	0.764 ± 0.032	0.651 ± 0.023	0.742 ± 0.062	4.36	0.0159
CHB	0.324 ± 0.029	0.226 ± 0.008	0.256 ± 0.026	4.52	0.0137
<b>Pigments</b>					
Anthocyanins	1.5 ± 0.2	1.1 ± 0.2	1.3 ± 0.1	NS	
$A_{526 \text{ diff}} \text{ g}^{-1} \text{ fwt}$					
Anthocyanins/TChl.	3.5 ± 0.5	3.3 ± 0.4	3.3 ± 0.3	NS	
$A_{526 \text{ diff}} \text{ mol}^{-1}$					
UV absorbing pigments	213 ± 37	132 ± 20	104 ± 17	4.29	0.0224
Mean $A_{(320-280\text{nm})} \text{ g}^{-1} \text{ dwt}$					
Total Chlorophyll	500 ± 58	374 ± 49	393 ± 39	NS	
$\text{nmol g}^{-1} \text{ fwt}$					
Chl a:b ratio	3.2 ± 0.2	3.3 ± 0.1	3.4 ± 0.2	NS	
VAZ/TChl	124 ± 14	93 ± 6	61 ± 7	4.72	0.0114
$\text{mmol mol}^{-1}$					
%(AZ/VAZ)	29.7 ± 3.0	40.4 ± 3.0	28.5 ± 6.3	3.68	0.0292
Tcar./Tchl.	677 ± 55	586 ± 29	458 ± 16	3.19	0.0462
$\text{mmol mol}^{-1}$					

Table 6. Reflectance signals of moss over all sites and species, and their Spearman rank correlation (*Rho*) and linear relationship with chlorophyll concentrations. Reflectance signals are from key wavelengths over the spectra measured. Regressions were done on logarithmically transformed data where appropriate. Levels of significance are \* $P < 0.10$ , \*\*  $P < 0.05$ , \*\*\*  $P < 0.001$ .

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Reflectance Parameter	<i>Rho</i>	Regression	$r^2$
R280	0.087	NS	
R320	0.057	NS	
R476	0.044	NS	
R526	0.298	LogR526 = -1.72 + 0.000193 x TChl	0.060**
R550	0.362	LogR550 = -1.60 + 0.000242 x TChl	0.080***
R680	-0.168	NS	
R850	0.486	R850 = 0.261 + 0.000134 x TChl	0.166***
R850-280	0.484	R850-280 = 0.254 + 0.000133 x TChl	0.161***
R850-320	0.482	R850-320 = 0.253 + 0.000133 x TChl	0.163***
R850-480	0.485	R850-480 = 0.251 + 0.000134 x TChl	0.166***
R850-526	0.475	R850-526 = 0.239 + 0.000125 x TChl	0.161***
R850-550	0.483	R850-550 = 0.259 + 0.000125 x TChl	0.171***
R850-680	0.488	R850-680 = 0.239 + 0.000132 x TChl	0.179***
R850/480	0.292	R850/480 = 31.83 + 0.0140 x TChl	0.050**
R850/526	0.063	NS	
R850/550	-0.007	NS	
R850/680	0.462	R850/550 = 13.46 + 0.0139 x TChl	0.107***
R680/850	-0.498	R680/850 = 0.103 + 0.00000651 x TChl	0.122***
ARE	0.335	$R_{ARE} = 698.8 + 0.00139 \times TChl$	0.069**
RE	0.488	$R_{RE} = 0.00562 + 0.00000365 \times TChl$	0.147***
PRI	0.273	$PRI = -0.194 + 0.00000596 \times TChl$	0.059**
NDVI	0.498	$NDVI = 0.813 + 0.000113 \times TChl$	0.125***
EGFR	0.196	NS	
EGFN	0.107	NS	
NPCI	-0.146	NS	
CHB	0.408	$CHB = 0.198 + 0.000162 \times TChl$	0.151***

Table 7. Reflectance of the moss *Grimmia antarctici* when wet and dry. Samples are from ridges ( $N=4$ ). Tests are paired t-tests for samples that were dry and then rehydrated for 5 minutes. NS signifies a non-significant difference at the  $P=0.10$  level).

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	Dry	Wet	<i>t</i>	<i>P</i>
R320	$0.0093 \pm 0.0014$	$0.0104 \pm 0.0013$	NS	
R526	$0.026 \pm 0.004$	$0.0022 \pm 0.004$	-2.62	0.079
R550	$0.031 \pm 0.010$	$0.037 \pm 0.004$	NS	
R850	$0.302 \pm 0.020$	$0.294 \pm 0.021$	NS	
RE	$0.0052 \pm 0.0001$	$0.109 \pm 0.005$	-21.12	0.0002
$\lambda_{RE}$	$713.25 \pm 0.75$	$699.9 \pm 0.41$	15.63	0.006
PRI	$-0.146 \pm 0.042$	$-0.118 \pm 0.046$	-4.87	0.016
NDVI	$0.881 \pm 0.020$	$0.890 \pm 0.018$	NS	
CHB	$0.287 \pm 0.021$	$0.260 \pm 0.031$	2.92	0.062

Table 8. Summary of effects of reflectance parameters and pigments over species, sites and microtopography. X indicates a main effect of the factor at  $P < 0.10$ , XX at  $P < 0.05$  and XXX at  $P < 0.001$ . N/A signifies not applicable because measurements were not made.

Parameter	Species	Site	Microtopography	Water Content
<b>Reflectance</b>				
R320				
R526	XXX	XX		X
R550	XX	XX		
R850	XX	XX		
RE	XXX	XX		XXX
$\lambda$ RE	XX	XXX		XX
PRI			XXX	XX
NDVI				
NPCI				
EGFR		XX		
EGFN		XX		
CHB	X	XX		X
<b>Pigments</b>				
Anthocyanins				N/A
$A_{526 \text{ diff}} \text{ g}^{-1} \text{ fwt}$				
Anthocyanins/TChl.	XX			N/A
$A_{526 \text{ diff}} \text{ mol}^{-1}$				
UV absorbing pigments	XXX	XX	N/A	N/A
Mean $A_{(320-280\text{nm})} \text{ g}^{-1} \text{ dwt}$				
Total Chlorophyll	X		XXX	N/A
$\text{nmol g}^{-1} \text{ fwt}$				
Chl a:b ratio				N/A
VAZ/TChl	XX	XX	XX	N/A
$\text{mmol mol}^{-1}$				
%(AZ/VAZ)	XX	XX	XX	N/A
Tcar./Tchl.	X	XX	XX	N/A
$\text{mmol mol}^{-1}$				

## Figure Captions

Figure 1. Mean reflectance spectra (A) and first derivative (B) of all samples of the three  
 5 common Antarctic moss species: *Bryum pseudotriquetrum* (dotted line, N=3), *Ceratodon  
 purpureus* (dashed line, N=5) and *Grimmia antarctici* (solid line, N=25) collected during the  
 1997/8 season from the Robinson Ridge and Red shed sites.

Figure 2. Relationships between total chlorophyll concentration of moss and the reflectance at  
 10 850 nm (A), amplitude of the red edge,  $\lambda_{RE}$  (B), and the NDVI (C). Lines of best fit are  
 exponential curves:  $R_{850} = 0.358 + [1 - \exp(-0.0099 \times TChl)]$ ,  $r^2 = 0.352$ ;  $\lambda_{RE} = 0.0084 + [1 - \exp(-$   
 $0.0079 \times TChl)]$ ,  $r^2 = 0.308$ .

Figure 3. Relationships between the Photochemical Reflectance Index (PRI) and the proportion  
 15 of the xanthophyll-cycle pigments in their active form ( $\%A+Z/V+A+Z$ ) (A) and the chlorophyll  
 fluorescence parameter  $F_v/F_m$  (B). Regression equations are:  $PRI = -0.0976 + (-0.00209 \times$   
 $\%AZ/VAZ)$ ,  $r^2 = 0.298$ ; and  $PRI = -1.285 + (1.604 \times F_v/F_m)$ ,  $r^2 = 0.427$ .





